

A review of natural products with antileishmanial activity

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Abstract

Infections caused by protozoa of the genus *Leishmania* are a major worldwide health problem, with high endemicity in developing countries. The incidence of the disease has increased since the emergence of AIDS. In the absence of a vaccine, there is an urgent need for effective drugs to replace/supplement those in current use. The plant kingdom is undoubtedly valuable as a source of new medicinal agents. The present work constitutes a review of the literature on plant extracts and chemically defined molecules of natural origin showing antileishmanial activity. The review refers to 101 plants, their families, and geographical distribution, the parts utilized, the type of extract and the organism tested. It also includes 288 compounds isolated from higher plants and microorganisms, classified into appropriate chemical groups. Some aspects of recent antileishmanial-activity-directed research on natural products are discussed.

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Keywords: *Leishmania*; Antileishmaniasis; Antileishmanial activity; Leishmanicidal activity; Medicinal plants; Natural products

Introduction

Leishmaniasis is regarded as a major public health problem (WHO, 2002), causing significant morbidity and mortality in Africa, Asia and Latin America. The disease currently threatens about 350 million women, men and children in 88 countries around the world, with about 2 million affected annually. In Brazil, studies report the occurrence of about 20,000 new cases of the illness annually. An increase in the incidence of leishmaniasis can be associated with urban development, forest devastation, environmental changes and migrations of people to areas where the disease is endemic (Carvalho et al., 2000; Patz et al., 2000; Ashford, 2000).

Species of the genus *Leishmania*, a protozoan member of the hemoflagellate group, are the causative agents of human leishmaniasis, which has a reservoir in rodents, dogs, saguins, marsupials and others in the wild animal population, and is transmitted by mosquitoes of the genera *Lutzomia* and *Phlebotomus*. The term leishmaniasis comprises three clearly distinguishable clinical manifestations: generalized visceral infection (visceral leishmaniasis or “Kala-azar”), cutaneous leishmaniasis (Oriental button), and mucocutaneous leishmaniasis (ulceration of the skin and hyperdevelopment of the mucous membranes) (Garcia-Granados et al., 1997; Ashford, 2000).

Members of the genus *Leishmania* differentiate from proliferative promastigotes in the sandfly vector gut to infective metacyclic promastigotes in the insect foregut. Parasites are inoculated by the vector as the flagellate promastigotes enter the mammalian host, where they infect macrophages, differentiating into nonmotile

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amastigotes and multiplying as such (Araújo et al., 1998; Carvalho et al., 2000). The mechanisms by which visceral and cutaneous manifestations develop have not been fully clarified.

The treatment of leishmaniasis is difficult because of the intramacrophagic location of the infectious form. Victims of this illness present an immune deficiency and are not able to eliminate the parasites through a natural mechanism of defense. Moreover, malnutrition is associated with certain cases of leishmaniasis. Parallel infection with diseases such as malaria and pneumonia increases the fatality of the illness if it is not diagnosed and treated in time. The problem of leishmaniasis has been worsened by the evolution of AIDS due to parallel infections in AIDS patients, as well as by the development of drug-resistance by parasites (Carvalho et al., 2000; Torres-Santos et al., 1999).

In the absence of a vaccine, there is an urgent need for effective drugs to replace/supplement those in current use. The clinically used drugs, many of which are based on pentavalent antimony compounds, were developed before 1959. The toxicity of these agents and the persistence of side-effects even after modification of the dose level and duration of treatment are, however, severe drawbacks. The search for antileishmanial agents has been exhaustive. Alternative drugs, such as amphotericin B and pentamidine, also have unpleasant side-effects (Balana et al., 1998; Carvalho et al., 2000). On the other hand, plant extracts or plant-derived compounds are likely to provide a valuable source of new medicinal agents (Carvalho and Ferreira, 2001; Kayser and Kiderlen, 2001) and the urgent need for alternative treatments has led to a program to screen natural products for potential use in the therapy of leishmaniasis. In fact, the WHO advocated the use of traditional medicine where appropriate health services are inaccessible (Tahir et al., 1998; Weniger et al., 2001; Bhadra, 1993).

Furthermore, the leads obtained from the search for natural products with antileishmanial activity give new impetus for obtaining valuable synthetic compounds (Carvalho et al., 2000).

With the objective of contributing to these studies, a literature search on the use of natural products (crude plant extracts, semi-purified fractions and chemically defined molecules) which have already been evaluated particularly for leishmaniasis, has been carried out.

Materials and methods

The keywords used for the literature search for this review were *Leishmania* × antileishmaniasis × antileishmanial activity × leishmanicidal activity × medicinal plants × natural products. The search was

carried out using Biological Abstracts, Chemical Abstracts, and the data bank of the University of Illinois in Chicago NAPRALERT (Acronym for NATural PRoducts ALERT), updated to December 2001. The references found in the search were then studied in detail.

Results and discussion

Consultation of various literature sources resulted in the elaboration of a list of natural products evaluated for antileishmanial activity (Tables 1 and 2). It should be noted that most of the references cited are not first-hand observations, but compilations copied from other sources. For details on the models or mechanism-based bioassays utilized for selecting crude plant extracts, fractions and pure compounds against the *Leishmania* parasite, the original references should be consulted.

Antileishmanial activity of crude plant extracts and fractions

A plant-screening program for potential leishmanicides was initiated in 1984 in French Guiana, based on the ethnomedical knowledge of the local population. The leishmanicidal activity of several plant extracts was evaluated *in vitro*, by testing on amastigote stages of *Leishmania amazonensis*, and *in vivo* using cutaneous *L. amazonensis* lesions in mice. Among the selected species, *Faramaea guianensis* showed activity (Sauvain et al., 1994).

Fourteen plants used topically in folk medicine in Bolivia to treat cutaneous leishmaniasis were collected in the tropical regions of colonization and in the rain forest occupied by Chimane Indians. Two plants employed by those in the colonial region showed an *in vitro* antileishmanial activity: *Bocconia integrifolia* and *B. pearcei*. Three other plants, *Ampelocera edentula*, *Galipea longiflora* and *Pera benensis*, employed by Chimane Indians, were effective in mice infected with *L. amazonensis* (Fournet et al., 1994b).

A preliminary examination of the crude methanol extracts of eight plant species collected from the Sudan revealed that only three plant species had a considerable *in vitro* antileishmanial activity on *L. major* promastigotes at a concentration <0.5 µg/ml. The plants *Azadirachta indica*, *Maytenus senegalensis* and *Eucalyptus globulus* showed IC₅₀ values of 11.5, 55 and 78 µg/ml, respectively (Tahir et al., 1998).

Singha et al. (1992) evaluated a total of 23 plants from Madras, India, for antileishmanial activity, with *L. donovani* infected hamsters. Extracts derived from five plants (viz., *Alstonia scholaris*, *Swertia chirata*, *Tibouchina semidecandra*, *Tinospora cordifolia* and *Nyctanthes*

Table 1. Plant extracts summary showing antileishmanial activity^a

Family and botanical name	Origin	Part used	Organism tested	Preparation ^b	Reference
Agavaceae					
<i>Yucca filamentosa</i> L.	Germany	^c	<i>L. amazonensis</i>	Ethanol extract	Plock et al. (2001)
Annonaceae					
<i>Annona glauca</i> Thonn.	Senegal	SD	<i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. donovani</i>	Dichloromethane ext Dichloromethane ext Dichloromethane ext	Waechter et al. (1998) Waechter et al. (1998) Waechter et al. (1998)
<i>Annona muricata</i> L.	Colombia	PE	<i>L. braziliensis</i> <i>L. panamensis</i>	Ethyl acetate extract Ethyl acetate extract	Jaramillo et al. (2000) Jaramillo et al. (2000)
<i>Annona senegalensis</i> Pers	Senegal	SD	<i>Leishmania</i> sp.	Dichloromethane ext	Sahpaz et al. (1996)
<i>Annona aff. spraguei</i> Saff.	Colombia	SD	<i>L. braziliensis</i> <i>L. infantum</i> <i>L. panamensis</i>	Chloroform extract Chloroform extract Chloroform extract	Saez et al. (1998) Saez et al. (1998) Saez et al. (1998)
<i>Cardiopetalum calophyllum</i> Schldl.	Bolivia	LF + SM	<i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. donovani</i>	Alkaloid fraction Alkaloid fraction Alkaloid fraction	Fournet et al. (1994b) Fournet et al. (1994b) Fournet et al. (1994b)
<i>Duguetia spixiana</i> Mart.	Bolivia	SB	<i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. donovani</i>	Alkaloid fraction Alkaloid fraction Alkaloid fraction	Fournet et al. (1994b) Fournet et al. (1994b) Fournet et al. (1994b)
<i>Guatteria foliosa</i> Benth	Bolivia	SB	<i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. donovani</i>	Alkaloid fraction Alkaloid fraction Alkaloid fraction	Mahiou et al. (1994) Mahiou et al. (1994) Mahiou et al. (1994)
<i>Guatteria schoburgkiana</i> Mart.	Bolivia	RB	<i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. donovani</i>	Alkaloid fraction Alkaloid fraction Alkaloid fraction	Fournet et al. (1994b) Fournet et al. (1994b) Fournet et al. (1994b)
<i>Oxandra espintana</i> (Spruce) Baillon	Bolivia	SB	<i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. donovani</i>	Alkaloid fraction Alkaloid fraction Alkaloid fraction	Fournet et al. (1994b) Fournet et al. (1994b) Fournet et al. (1994b)
<i>Xylopia aromatica</i> (Lam.) Mart.	Bolivia	LF + SB	<i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. donovani</i>	Alkaloid fraction Alkaloid fraction Alkaloid fraction	Fournet et al. (1994b) Fournet et al. (1994b) Fournet et al. (1994b)
Apocynaceae					
<i>Alstonia scholaris</i> R. Br.	India	SM	<i>L. donovani</i>	Ethanol extract	Singha et al. (1992)
<i>Holarrhena curtisii</i> King & Gamble	Malaysia	LF	<i>L. donovani</i>	Ethanol extract	Kam et al. (1998)
<i>Mandevilla antennacea</i> K. Schum.	Bolivia	LF + SM	<i>L. amazonensis</i> <i>L. braziliensis</i>	Ethanol extract Ethanol extract	Fournet et al. (1994b) Fournet et al. (1994b)
<i>Peschiera australis</i> Miers.	Brazil	SM	<i>L. amazonensis</i>	Chloroform extract	Delorenzi et al. (2001)
<i>Peschiera</i> var. <i>heurkii</i> (Muell. Arg.) L. Allorge	Bolivia	LF	<i>L. amazonensis</i> <i>L. braziliensis</i>	Ethanol extract Ethanol extract	Munoz et al. (1994) Munoz et al. (1994)
<i>Picralima nitida</i> Th. & H. Dur.	Nigeria	SD	<i>L. donovani</i>	Chloroform extract	Iwu et al. (1992)
<i>Tabernaemontana obliqua</i> (Miers) Leenwenb.	Colombia	LF	<i>L. amazonensis</i>	Methanol extract	Weniger et al. (2001)
Araliaceae					
<i>Hedera helix</i> L.	Spain	LF	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)

<i>Oreopanax</i> species Asclepiadaceae	Bolivia	LF	<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
<i>Gongronema latifolia</i> Benth	Nigeria	LF	<i>L. donovani</i>	Methanol extract	Iwu et al. (1992)
<i>Periploca graeca</i> L.	Turkey	TG	<i>L. major</i>	Methanol extract	Demirci et al. (1998)
Asteraceae					
<i>Acanthospermum hispidum</i> DC.	Bolivia	EP	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
<i>Achyrocline flaccida</i> (Weinm.) DC.	Bolivia	EP	<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
<i>Ageratina pentlandiana</i> (DC.) K. & R.	Bolivia	LF	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethanol extract.	Lockman et al. (1991)
<i>Artemisia herba-alba</i> Asso.	Maroc	c	<i>L. tropica</i>	Aqueous extract	Hatimi et al. (2001)
<i>Baccharis salicifolia</i> (R. & P.) Pers.	Bolivia	LF	<i>L. braziliensis</i>	Ethyl acetate extract	Fournet et al. (1994b)
<i>Chersodoma jodopappa</i> (Sch. Bip.) Cabrera	Bolivia	LF + SM	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. donovani</i>	Ethanol extract	Fournet et al. (1994b)
<i>Cnicothamnus lorentzii</i> Griseb.	Bolivia	LF + SM	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. donovani</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
<i>Echinacea purpurea</i> Moench	Germany	EP	<i>Leishmania</i> sp.	Sap	Parnham (1996)
<i>Inula montana</i> L.	Spain	AP	<i>L. infantum</i>	Methanol	Martin et al. (1998)
<i>Jasonia glutinosa</i> DC.	Spain	AP	<i>L. donovani</i>	Acetone extract	Villaescusa et al. (1996)
<i>Munozia fournetii</i> H. Robinson	Bolivia	LF + SM	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. donovani</i>	Ethanol extract	Fournet et al. (1994b)
<i>Neurolaena lobata</i> R. Br.	Guatemala	LF	<i>L. mexicana</i>	Ethanol extract	Berger et al. (2001)
			<i>L. braziliensis</i>	Ethanol extra	Berger et al. (2001)
<i>Ophryosporus piquerioides</i> (DC.) Benth.	Bolivia	EP	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. donovani</i>	Ethanol extract	Fournet et al. (1994b)
<i>Perezia multiflora</i> Less. (H. & B.) Less.	Bolivia	LF	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. donovani</i>	Ethanol extract	Fournet et al. (1994b)
<i>Pterocaulon alopecuroideum</i> (Lam.)DC.	Bolivia	EP	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. donovani</i>	Ethanol extract	Fournet et al. (1994b)
<i>Senecio clivicolus</i> Wedd.	Bolivia	LF + SM	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. donovani</i>	Ethanol extract	Fournet et al. (1994b)
<i>Stevia yaconensis</i> Hieron.	Bolivia	EP	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. donovani</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
<i>Vernonia squamulosa</i> Hook. & Arn.	Bolivia	SM	<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. donovani</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
<i>Werneria nubigena</i> H.B.K.	Bolivia	LF + SM	<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)

Table 1. (continued)

Family and botanical name	Origin	Part used	Organism tested	Preparation ^b	Reference
<i>Xanthium catharticum</i> L.	Bolivia	RT + SM	<i>L. donovani</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. donovani</i>	Ethanol extract	Fournet et al. (1994b)
			<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
Berberidaceae					
<i>Berberis boliviana</i> Lechl.	Bolivia	BK + SM	<i>L. amazonensis</i>	Alkaloid fraction	Fournet et al. (1994b)
<i>Berberis bumeliaefolia</i> Schum	Bolivia	BK	<i>L. braziliensis</i>	Alkaloid fraction	Fournet et al. (1994b)
			<i>L. donovani</i>	Alkaloid fraction	Fournet et al. (1994b)
			<i>L. amazonensis</i>	Alkaloid fraction	Fournet et al. (1994b)
<i>Berberis cf. laurina</i> Epl.	Bolivia	SM	<i>L. braziliensis</i>	Alkaloid fraction	Fournet et al. (1994b)
			<i>L. donovani</i>	Alkaloid fraction	Fournet et al. (1994b)
			<i>L. amazonensis</i>	Alkaloid fraction	Fournet et al. (1994b)
<i>Berberis</i> aff. <i>paucidentata</i> Rusby	Bolivia	SB	<i>L. donovani</i>	Alkaloid fraction	Fournet et al. (1994b)
			<i>L. amazonensis</i>	Alkaloid fraction	Fournet et al. (1994b)
Bignoniaceae					
<i>Jacaranda copaia</i> D. Don	Fr. Guiana	LF	<i>L. amazonensis</i>	Type extract not stated	Sauvain et al. (1993)
<i>Kigelia pinnata</i> DC.	Africa	RB	<i>L. major</i>	Type extract not stated	Moideen et al. (1997)
Bombacaceae					
<i>Huberodendron patinoi</i> Cuatrec.	Colombia	BK	<i>L. panamensis</i>	Methanol extract	Weniger et al. (2001)
Burseraceae					
<i>Protium amplum</i> Cuatrec.	Colombia	FR	<i>L. amazonensis</i>	Methylene chloride extract	Weniger et al. (2001)
			<i>L. braziliensis</i>	Methylene chloride extract	Weniger et al. (2001)
			<i>L. infantum</i>	Methylene chloride extract	Weniger et al. (2001)
Caparraceae					
<i>Capparis spinosa</i> L.	Israel	BC	<i>L. major</i>	Type extract not stated	Schlein (1994)
Celastraceae					
<i>Maytenus senegalensis</i> (Lam.) Exell	Sudan	SB	<i>L. major</i>	Dichloromethane ext	Tahir et al. (1998)
Clusiaceae					
<i>Marila laxiflora</i> Rusby	Colombia	LF	<i>L. amazonensis</i>	Methylene chloride extract	Weniger et al. (2001)
			<i>L. braziliensis</i>	Methylene chloride extract	Weniger et al. (2001)
Crassulaceae					
<i>Bryophyllum pinnatum</i> Kurz	Brazil	LF	<i>L. amazonensis</i>	Aqueous extract Aqueous extract	Da Silva et al. (1995) Rossi et al. (2000)
Dilleniaceae					
<i>Doloiocarpus dentatus</i> Kubitzki	Surinam	SM	<i>L. amazonensis</i>	Chloroform extract	Sauvain et al. (1996)

Euphorbiaceae					
<i>Pera benensis</i> Rusby	Bolivia	RB + SB	<i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. donovani</i>	Ethanol extract Ethanol extract Ethanol extract	Fournet et al. (1994b) Fournet et al. (1994b) Fournet et al. (1994b)
<i>Ricinus communis</i> V.A. Moshkin	Israel	BC	<i>L. major</i>	Type extract not stated	Schlein (1994)
Fabaceae					
<i>Crotalaria barbata</i> R. Grah.	India	EP	<i>L. donovani</i>	Ethanol extract	Singha et al. (1992)
<i>Desmodium gangeticum</i> L.	Nigeria	LF	<i>L. donovani</i>	Methanol extract	Iwu et al. (1992)
<i>Periandra mediterranea</i> Taub.	Brazil	RT	<i>L. donovani</i>	Saponin fraction	Santo et al. (1997)
<i>Spartium junceum</i> L.	Israel	BC	<i>L. major</i>	Type extract not stated	Schlein (1994)
Gentianaceae					
<i>Swertia chirata</i> Buch. Ham. Ex Wall.	India	EP	<i>L. donovani</i>	Ethanol extract	Singha et al. (1992)
Geraniaceae					
<i>Pelargonium sidoides</i> DC.	Germany	°	<i>L. donovani</i>	Ethanol extract	Kayser et al. (2001a)
Lauraceae					
<i>Aniba canelilla</i> H.B.K.	Bolivia	SM	<i>L. amazonensis</i> <i>L. braziliensis</i>	Ethyl acetate extract Ethyl acetate extract	Fournet et al. (1994b) Fournet et al. (1994b)
<i>Aniba</i> species	Bolivia	SM	<i>L. amazonensis</i> <i>L. braziliensis</i>	Ethanol extract Ethanol extract	Fournet et al. (1994b) Fournet et al. (1994b)
Liliaceae					
<i>Allium sativum</i> L.	Iran	BULB	<i>L. major</i>	°	Ghazanfari et al. (2000)
Malvaceae					
<i>Malva nicaeensis</i> All.	Israel	BC	<i>L. major</i>	Type extract not stated	Schlein (1994)
Melastomaceae					
<i>Tibouchina semidecandra</i> Cogn.	India	AP	<i>L. donovani</i>	Ethanol extract	Singha et al. (1992)
Meliaceae					
<i>Azadirachta indica</i> A. Juss.	Sudan	SB	<i>L. major</i>	Methanol extract	Tahir et al. (1998)
<i>Guarea polymera</i> Little	Colombia	LF	<i>L. amazonensis</i> <i>L. braziliensis</i>	Methylene chloride extract Methylene chloride extract	Weniger et al. (2001) Weniger et al. (2001)
<i>Khaya senegalensis</i> A. Juss.	Portugal	°	<i>L. donovani</i>	°	Abreu et al. (1999)
Menispermaceae					
<i>Abuta pahnii</i> Mart.	Bolivia	SM	<i>L. amazonensis</i> <i>L. braziliensis</i>	Alkaloid fraction Alkaloid fraction	Fournet et al. (1994b) Fournet et al. (1994b)
<i>Abuta rufescens</i> Aublet	Bolivia	BK	<i>L. amazonensis</i>	Alkaloid fraction	Fournet et al. (1994b)
<i>Anomospermum bolivianum</i> Kruk. & Mold	Bolivia	BK	<i>L. amazonensis</i> <i>L. braziliensis</i>	Alkaloid fraction Alkaloid fraction	Fournet et al. (1994b) Fournet et al. (1994b)
<i>Tinospora cordifolia</i> (Willd.) Hook.f. & Thoms.	India	SM	<i>L. donovani</i>	Ethanol extract	Singha et al. (1992)
Moraceae					
<i>Dorstenia multiradiata</i> Engl.	Nigeria	LF	<i>L. donovani</i>	Aqueous extract	Iwu et al. (1992)
Myristicaceae					
<i>Otoba novogranatensis</i> Moldenke	Colombia	LF	<i>L. amazonensis</i>	Methylene chloride extract	Weniger et al. (2001)

Table 1. (continued)

Family and botanical name	Origin	Part used	Organism tested	Preparation ^b	Reference
			<i>L. braziliensis</i>	Methylene chloride extract	Weniger et al. (2001)
			<i>L. infantum</i>	Methylene chloride extract	Weniger et al. (2001)
			<i>L. amazonensis</i>	Methanol extract	Weniger et al. (2001)
			<i>L. braziliensis</i>	Methanol extract	Weniger et al. (2001)
			<i>L. infantum</i>	Methanol extract	Weniger et al. (2001)
<i>Otoba parvifolia</i> (Mgf.) A.H. Gentry	Colombia	BK	<i>L. amazonensis</i>	Methylene chloride extract	Weniger et al. (2001)
			<i>L. braziliensis</i>	Methylene chloride extract	Weniger et al. (2001)
Myrsinaceae					
<i>Myrsine pellucida</i> Spreng	Bolivia	SB	<i>L. braziliensis</i>	Ethanol extract	Lavaud et al. (1994)
Papaveraceae					
<i>Bocconia integrifolia</i> H and B	Bolivia	LF + SB	<i>L. amazonensis</i>	Alkaloid fraction	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Alkaloid fraction	Fournet et al. (1994b)
			<i>L. donovani</i>	Alkaloid fraction	Fournet et al. (1994b)
<i>Bocconia pearcei</i> Hutch.	Bolivia	LF	<i>L. amazonensis</i>	Alkaloid fraction	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Alkaloid fraction	Fournet et al. (1994b)
			<i>L. donovani</i>	Alkaloid fraction	Fournet et al. (1994b)
Phytolaccaceae					
<i>Phytolacca dodecandra</i> L'Herit.	Ethiopia	FR	<i>L. enriettii</i>	Buthanol extract	Lemma et al. (1972)
Piperaceae					
<i>Peperomia galioides</i> H. B. & K.	Bolivia	EP	<i>L. amazonensis</i>	Ethanol extract	
			<i>L. braziliensis</i>	Ethanol extract	Mahiou et al. (1995)
			<i>L. chagasi</i>	Ethanol extract	Mahiou et al. (1995)
			<i>L. donovani</i>	Petroleum ether extract	Mahiou et al. (1995)
<i>Piper aduncum</i> L.	Brazil	IF	<i>L. amazonensis</i>	Petroleum ether extract	Torres-Santos et al. (1999)
<i>Piper rusbyi</i> C. DC	Bolivia	EP	<i>L. amazonensis</i>	Ethyl acetate extract	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Ethyl acetate extract	Fournet et al. (1994b)
			<i>L. donovani</i>	Ethyl acetate extract	Fournet et al. (1994b)
Rubiaceae					
<i>Faramea guianensis</i> (Aubl.) Bremek	Fr Guiana	LF	<i>L. amazonensis</i>	Aqueous extract	Sauvain et al. (1994)
Rutaceae					
<i>Dictyoloma peruvianum</i> Planch.	Bolivia	SB	<i>L. amazonensis</i>	Ethyl acetate extract	Lavaud et al. (1995)
			<i>L. braziliensis</i>	Alkaloid fraction	Lavaud et al. (1995)
<i>Galipea longiflora</i> Kr	Bolivia	LF	<i>L. amazonensis</i>	Alkaloid fraction	Fournet et al. (1994a)
			<i>L. braziliensis</i>	Alkaloid fraction	Fournet et al. (1994a)
			<i>L. donovani</i>	Alkaloid fraction	Fournet et al. (1994a)
		LF + RB	<i>L. amazonensis</i>	Alkaloid fraction	Fournet et al. (1994b)
			<i>L. braziliensis</i>	Alkaloid fraction	Fournet et al. (1994b)
			<i>L. donovani</i>	Alkaloid fraction	Fournet et al. (1994b)
		RB	<i>L. amazonensis</i>	Alkaloid fraction	Fournet et al. (1994a)
			<i>L. braziliensis</i>	Alkaloid fraction	Fournet et al. (1994a)

<i>Swinglea glutinosa</i> Merr.	Colombia	BK	<i>L. donovani</i> <i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. infantum</i>	Alkaloid fraction Methylene chloride extract Methylene chloride extract Methylene chloride extract	Fournet et al. (1993a) Weniger et al. (2001) Weniger et al. (2001) Weniger et al. (2001)
Sapindaceae					
<i>Serjania tenuifolia</i> Radlk	Bolivia	LF + SM	<i>L. amazonensis</i> <i>L. braziliensis</i>	Ethanol extract Ethanol extract	Fournet et al. (1994b) Fournet et al. (1994b)
Scrophulariaceae					
<i>Conohea scoparioides</i> (Cham. & Schltdl.) Benth	Colombia	LF	<i>L. amazonensis</i> <i>L. braziliensis</i>	Methylene chloride extract Methylene chloride extract	Weniger et al. (2001) Weniger et al. (2001)
<i>Picrorhiza kurroa</i> Royle, ex Benth	India	RZ + RT	<i>L. donovani</i>	Ethanol extract	Mittal et al. (1998)
<i>Scrophularia scorodonia</i> L.	Spain	FL	<i>L. infantum</i>	Methanol extract	Martin et al. (1998)
Solanaceae					
<i>Nicotiana glauca</i> Graham.	Israel	LF + SM	<i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. donovani</i>	Ethanol extract Ethanol extract Ethanol extract	Fournet et al. (1994b) Fournet et al. (1994b) Fournet et al. (1994b)
<i>Saracha punctata</i> Ruiz & Pav.	Bolivia	LF	<i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. donovani</i>	Ethanol extract Ethanol extract Ethanol extract	Moretii et al. (1998) Moretii et al. (1998) Moretii et al. (1998)
<i>Solanum actaeabotrys</i> Rusby	Bolivia	LF	<i>L. amazonensis</i> <i>L. braziliensis</i> <i>L. donovani</i>	Ethanol extract Ethanol extract Ethanol extract	Fournet et al. (1994b) Fournet et al. (1994b) Fournet et al. (1994b)
<i>Solanum luteum</i> Mill	Israel	BC	<i>L. donovani</i> <i>L. major</i>	Ethanol extract Type extract not stated	Fournet et al. (1994b) Schlein (1994)
Sterculiaceae					
<i>Cola attiensis</i> Aubrev. & Pellegr.	Nigeria	SD	<i>L. donovani</i>	Chloroform extract	Iwu et al. (1992)
Ulmaceae					
<i>Ampelocera edentula</i> Kulm	Bolivia	SM	<i>L. amazonensis</i>	Ethanol extract	Fournet et al. (1994b)
Verbenaceae					
<i>Nyctanthes arbortristis</i> L.	India	AP	<i>L. donovani</i>	Ethanol extract	Rathore et al. (1989)
<i>Vitex heterophylla</i> Miq.	India	LF	<i>L. donovani</i>	Ethanol extract	Bhakuni et al. (1988)

^aOnly positive plant extract tested for antileishmanial activity were presented in Table 1. Data for negative results is available from the authors on request.

^bIn most cases the reference compound to a positive control were either glucontime or pentamidine.

^cDate incomplete derived from an abstract; AP, aerial part; BC, branches; BK, bark; EP, entire plant; FL, flowers; FR, fruits; IF, inflorescence; LF, leaves; PE, pericarp; RB, rootbark; RT, roots; RZ, rhizome; SB, stembark; SD, seeds; SM, stem; TG, twig.

Table 2. Chemically defined molecules with antileishmanial activity ^a

Chemical substance ^b	Class	Organism tested	References
Acivicin	Alkaloid	<i>L. donovani</i>	Mukherjee et al. (1990)
Agarofuran, beta dihydro: 6-beta-8-beta-diacetoxy-1-alpha-9-alpha-dibenzoyloxy-15-(2)-methyl-butyroly-oxy-2-alpha-hydroxy	Sesquiterpene	<i>L. tropica</i>	Perez-Victoria et al. (1999)
Ajmalicine	Alkaloid	<i>L. major</i>	Staerk et al. (2000)
Alizarin, 3-methyl	Quinoid	<i>L. major</i>	Sittie et al. (1999)
Allopurinol	Alkaloid	<i>L. donovani</i>	Werbel and Worth (1980)
Aloe emodin	Quinoid	<i>L. donovani</i>	Camacho et al. (2000a)
Amarogentin	Iridoid	<i>L. donovani</i>	Medda et al. (1999)
Amentoflavone	Flavonoid	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Amphotericin B	Miscellaneous lactone	<i>Leishmania</i> sp. <i>L. braziliensis</i> <i>L. braziliensis</i> <i>L. donovani</i> <i>Leishmania</i> sp.	Chance (1995) Chalcat et al. (1965) Furtado et al. (1960) Neal and Croft (1984) Lyubimova et al. (1979)
Ancistroealaine A	Alkaloid	<i>L. donovani</i>	Bringman et al. (2000)
Ancistroealaine B	Alkaloid	<i>L. donovani</i>	Bringman et al. (2000)
Annonacin	Miscellaneous lactone	<i>L. amazonensis</i>	Waechter et al. (1998)
Annonacin A	Miscellaneous lactone	<i>L. amazonensis</i>	Waechter et al. (1998)
Anonaine, (–)	Alkaloid	<i>L. amazonensis</i>	Queiroz et al. (1996)
Anophonin	Lignan	<i>L. major</i>	Moideen et al. (1997)
Anthraquinone-2-hydroxymethyl-3-hydroxy	Quinoid	<i>L. major</i>	Sittie et al. (1999)
Anthraquinone-2-carbaldehyde	Quinoid	<i>L. major</i>	Chan-Bacab and Pena-Rodriguez (2001)
Antioquine	Alkaloid	<i>L. amazonensis</i>	Fournet et al. (1993d)
Aphidicolin	Diterpene	<i>L. donovani</i>	Kayser et al. (2001b)
Arbortristoside A	Iridoid	<i>L. donovani</i>	Tandon et al. (1991)
Arbortristoside B	Iridoid	<i>L. donovani</i>	Tandon et al. (1991)
Arbortristoside C	Iridoid	<i>L. donovani</i>	Tandon et al. (1991)
Argentilactone	Miscellaneous lactone	<i>L. amazonensis</i>	Waechter et al. (1997)
Argentinine	Alkaloid	<i>L. donovani</i>	Mahiou et al. (1994)
Aristeromycin, (–)	Alkaloid	<i>L. donovani</i>	Hiraoka et al. (1986)
Artemether	Sesquiterpene	<i>L. major</i>	Yang and Liew (1993)
Artemisinin	Sesquiterpene	<i>L. major</i>	Yang and Liew (1993)
Benzofuran, 6-benzoyl-2-(oxomethylphenyl)-3-hydroxy	Oxygen heterocycle	<i>L. donovani</i>	Kayser and Kiderlen (1999)
Benzofuranone, 3(2H): 4,6-dibenzoyl-2-(phenyl-hydroxy-methyl)	Oxygen heterocycle	<i>L. donovani</i>	Kayser and Kiderlen (1999)
Benzofuranone, 3(2H): 6-hydroxy-2-(phenyl-methylene)	Oxygen heterocycle	<i>L. donovani</i>	Kayser and Kiderlen (1999)
Benzofuranone, 3(2H): 6-methoxy-2-(phenyl-hydroxy-methylene)	Oxygen heterocycle	<i>L. donovani</i>	Kayser and Kiderlen (1999)
Benzofuranone, 3(2H): 6-methoxy-2-(phenyl-methylene)	Oxygen heterocycle	<i>L. donovani</i>	Kayser and Kiderlen (1999)
Benzoxazolinone	Alkaloid	<i>L. donovani</i>	Carvalho and Ferreira (2001)
Benzoxazol-2(3H)-one	Alkaloid	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Berbamine	Alkaloid	<i>L. amazonensis</i>	Fournet et al. (1993d)
Berberine	Alkaloid	<i>L. donovani</i>	Vennerstrom et al. (1990)

Berberine, dihydro	Alkaloid	<i>L. mexicana</i>	Edward et al. (1995)
Berberine, oxy	Alkaloid	<i>L. donovani</i>	Vennerstrom et al. (1990)
Berberine, tetrahydro	Alkaloid	<i>L. donovani</i>	Vennerstrom et al. (1990)
Berberine, tetrahydro: <i>N</i> -oxyde	Alkaloid	<i>L. donovani</i>	Vennerstrom et al. (1990)
Betuletol	Flavonoid	<i>Leishmania</i> sp.	Morales et al. (2000)
Betulin aldehyde	Triterpene	<i>L. amazonensis</i>	Sauvain et al. (1996)
Brachycalxolide, 16,17-dihydro	Sesquiterpene	<i>L. major</i>	Oketch-Rabah et al. (1998)
Brachycoumarinone, 2-cyclo epoxide	Coumarin	<i>L. major</i>	Oketch-Rabah et al. (1997a)
Brachycoumarinone, 2-epicyclo epoxide	Coumarin	<i>L. major</i>	Oketch-Rabah et al. (1997a)
Bractein	Flavonoid	<i>L. donovani</i>	Kayser et al. (1999)
		<i>L. donovani</i>	Kayser and Kiderlen (1999)
Bracteoline	Alkaloid	<i>L. amazonensis</i>	Queiroz et al. (1996)
Bruceantin	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Brucein A, iso	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Brucein B, iso	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Buchtienine	Alkaloid	<i>L. donovani</i>	Kam et al. (1999)
Buddlejasaponin	Triterpene	<i>L. infantum</i>	Emam et al. (1995)
Camptothecin	Alkaloid	<i>L. donovani</i>	Bodley and Shapiro (1995)
		<i>L. donovani</i>	Bodley et al. (1995)
Chalcone, 2',6'-dihydroxy-4'-methoxy	Flavonoid	<i>L. amazonensis</i>	Torres-Santos et al. (1999)
Chaparrinone	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Chaparrinone, 15-beta-heptyl	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Chimanine A	Alkaloid	<i>L. amazonensis</i>	Fournet et al. (1993c)
		<i>L. braziliensis</i>	Fournet et al. (1993a)
		<i>L. donovani</i>	Fournet et al. (1994a)
Chimanine B	Alkaloid	<i>L. amazonensis</i>	Fournet et al. (1994a)
		<i>L. braziliensis</i>	Fournet et al. (1994a)
		<i>L. donovani</i>	Fournet et al. (1994a)
Chimanine D	Alkaloid	<i>L. amazonensis</i>	Fournet et al. (1993c)
		<i>L. braziliensis</i>	Fournet et al. (1993a)
		<i>L. donovani</i>	Fournet et al. (1994a)
		<i>Leishmania</i> sp.	Munos et al. (1994)
Colchiside 4	Triterpene	<i>L. infantum</i>	Mshvildadze et al. (2000)
Colchiside 7	Triterpene	<i>L. infantum</i>	Mshvildadze et al. (2000)
Concanavalin B	Proteid	<i>L. amazonensis</i>	Barral-Neto et al. (1996)
Conoduramine	Alkaloid	<i>L. amazonensis</i>	Munoz et al. (1994)
Conodurine	Alkaloid	<i>L. amazonensis</i>	Munoz et al. (1994)
Coreximine, (–)	Alkaloid	<i>L. amazonensis</i>	Mahiou et al. (1994)
Coronaridine	Alkaloid	<i>L. amazonensis</i>	Delorenzi et al. (2001)
Corydine	Alkaloid	<i>L. donovani</i>	Camacho et al. (2000b)
Corynantheidine	Alkaloid	<i>L. major</i>	Staerk et al. (2000)
Corynantheidine, dihydro	Alkaloid	<i>L. major</i>	Staerk et al. (2000)
Corynanthine	Alkaloid	<i>L. major</i>	Staerk et al. (2000)
Coumarin	Coumarin	<i>L. amazonensis</i>	Bravo et al. (1999)
Cryptofolione	Miscellaneous lactone	<i>L. amazonensis</i>	Schmeda-Hirschmann et al. (2001)
Curcumin	Benzenoid	<i>L. major</i>	Rasmussen et al. (2000a)
Curcumin, bisdemethoxy	Benzenoid	<i>L. major</i>	Rasmussen et al. (2000a)

Table 2. (continued)

Chemical substance ^b	Class	Organism tested	References
Curcumin, demethoxy	Benzenoid	<i>L. major</i>	Rasmussen et al. (2000a)
Cusparine	Alkaloid	<i>L. amazonensis</i>	Fournet et al. (1993c)
Damnacanthal	Quinoid	<i>L. major</i>	Sittie et al. (1999)
Damnacanthal, nor	Quinoid	<i>L. major</i>	Sittie et al. (1999)
Daphnandrine	Alkaloid	<i>L. braziliensis</i>	Chan-Bacab and Pena-Rodriguez (2001)
Decanoic acid	Lipid	<i>L. donovani</i>	Cunningham et al. (1972)
Dehydropinifolic acid 15-monomethyl ester	Diterpene	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Diallyl disulfide	Sulfur compound	<i>Leishmania</i> sp.	McClure et al. (1996)
Dicentrinone	Alkaloid	<i>L. donovani</i>	Camacho et al. (2000b)
Dictyolamide A	Alkaloid	<i>L. amazonensis</i>	Lavaud et al. (1995)
Diospyrin	Quinoid	<i>L. donovani</i>	Hazra et al. (1995)
Diospyrin, hydroxy	Quinoid	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Dodeca-4-enoic acid, 4,6-diethyl-3(<i>S</i>)-5(<i>R</i>)-epidioxy-8-methyl	Lipid	<i>L. mexicana</i>	Compagnone et al. (1998)
Dodeca- <i>cis</i> -2,4-dienoic acid, 3,6(<i>R</i>)-epoxy-4,6-8(<i>S</i>)-triethyl: methyl ester	Lipid	<i>L. mexicana</i>	Compagnone et al. (1998)
Dodeca- <i>cis</i> -2,4-dienoic acid, 4,6-diethyl-3,6(<i>R</i>)-epoxy-8(<i>S</i>)-methyl: methyl ester	Lipid	<i>L. mexicana</i>	Compagnone et al. (1998)
Domesticine, nor:	Alkaloid	<i>L. amazonensis</i>	Queiroz et al. (1996)
(<i>E</i>)-1-[2,4-Dihydroxy-3-(3-methylbut-2-enyl)-phenyl]-3-[4-hydroxy-3-(3-methylbut-2-enyl)phenyl]prop-2-en-1-one	Flavonoid	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Emetine, (–)	Alkaloid	<i>L. tropica</i>	Neal (1970)
Emetine, 2,3-dehydro	Alkaloid	<i>L. tropica</i>	Neal (1970)
Espintanol	Monoterpene	<i>L. amazonensis</i>	Hocquemiller et al. (1991)
Eudesm-4(14)-ene-5- α -11(<i>R</i>)-12-triol	Sesquiterpene	<i>L. donovani</i>	Villaescusa et al. (2000)
Fissinolide	Triterpene	<i>L. major</i>	Khalid et al. (1998)
Flavone, iso: 3',7-dihydroxy-4'-methoxy	Flavonoid	<i>L. amazonensis</i>	Araújo et al. (1998)
Formycin B	Alkaloid	<i>L. donovani</i>	Neal and Croft (1984)
		<i>L. donovani</i>	Carson and Chang (1981)
		<i>L. major</i>	Neal et al. (1985)
Furan, tetrahydro: 2-(5-(2-hydroxy-undecyl)-2-tetrahydrofuran-5-yl)-5-(14-(5-methyl-2-oxo-2,5-dihydro-3-furanyl)-1,5,12-trihydroxy-tetradecyl)	Oxygen heterocycle	<i>L. donovani</i>	Cavé et al. (1990)
Gabunine	Alkaloid	<i>L. amazonensis</i>	Munoz et al. (1994)
Germacratien 2,5-epoxy-2 β -hydroxy-8 α -(2-methylpropenoyloxy)-4(15),10(14),11(13)-12,6 α -olide	Sesquiterpene	<i>L. major</i>	Fuchino et al. (2001)
Glaucifilin	Miscellaneous lactone	<i>L. amazonensis</i>	Waechter et al. (1998)
Glaucanisin	Miscellaneous lactone	<i>L. amazonensis</i>	Waechter et al. (1998)
Glaucaruantine	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Glaucarubin	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Glaucarubinone	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Glaucarubol, 15- β -glucosyl	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Glucan	Carbohydrate	<i>L. donovani</i>	Cook et al. (1980)
Goniothalamycin	Miscellaneous lactone	<i>L. amazonensis</i>	Waechter et al. (1998)
Grifolic acid	Sesquiterpene	<i>L. amazonensis</i>	Mahiou et al. (1995)

Grifolin	Sesquiterpene	<i>L. amazonensis</i>	Mahiou et al. (1995)
Guattouregidine, iso	Alkaloid	<i>L. braziliensis</i>	Fournet et al. (1996)
Gyrocarpine	Alkaloid	<i>L. amazonensis</i>	Mahiou et al. (1994)
Harmaline	Alkaloid	<i>Leishmania</i> sp.	Fournet et al. (1993d)
Harmine	Alkaloid	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Hederacolchiside A	Triterpene	<i>L. infantum</i>	Kam et al. (1999)
Hederacolchiside A'	Triterpene	<i>L. infantum</i>	Mshvildadze et al. (2000)
Hederacolchiside A-1	Triterpene	<i>L. amazonensis</i>	Mshvildadze et al. (2000)
Hederagenin	Triterpene	<i>L. tropica</i>	Ridoux et al. (2001)
Hederin, alpha	Triterpene	<i>L. tropica</i>	Savornin et al. (1991)
Hederin, beta	Triterpene	<i>L. tropica</i>	Savornin et al. (1991)
Hederin, delta	Triterpene	<i>L. tropica</i>	Savornin et al. (1991)
Heptane, 3-hydroxy-1,7-bis-(4',4''-dihydro-xyphenyl)	Benzenoid	<i>L. amazonensis</i>	Savornin et al. (1991)
Holacurtine	Steroid	<i>L. donovani</i>	Araújo et al. (1998)
Holacurtine, N-demethyl	Steroid	<i>L. donovani</i>	Kam et al. (1998)
Holacurtine, 17-epi	Steroid	<i>L. donovani</i>	Kam et al. (1998)
Holacurtine, 17-epi-N-demethyl	Steroid	<i>L. donovani</i>	Kam et al. (1998)
Holacurtinol	Steroid	<i>L. donovani</i>	Kam et al. (1998)
Holamine	Steroid	<i>L. donovani</i>	Kam et al. (1998)
Holamine, 15-alpha-hydroxy	Steroid	<i>L. donovani</i>	Kam et al. (1998)
Hyaluronic acid	Carbohydrate	<i>L. donovani</i>	Kam et al. (1998)
Ibericin	Quinoid	<i>L. major</i>	Seneca et al. (1948)
Incomptin B	Sesquiterpene	<i>L. mexicana</i>	Sittie et al. (1999)
Jacaranone	Quinoid	<i>L. amazonensis</i>	Guerrero et al. (1995)
Jatrogrossidione	Diterpene	<i>L. chagasi</i>	Sauvain et al. (1993)
Jatrophone	Diterpene	<i>L. amazonensis</i>	Schmeda-Hirschmann et al. (1996)
Kutkoside	Iridoid	<i>L. donovani</i>	Schmeda-Hirschmann et al. (1996)
Kudtriol	Sesquiterpene	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Labda-8-trans-13-dien-15-oic acid, 18-carboxy: methyl ester (4 <i>S</i> ,9 <i>R</i> ,10 <i>R</i>)	Diterpene	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Labda-9-14-diene, 3-beta-acetoxy-8-alpha-13-epoxy-12-oxo	Diterpene	<i>L. donovani</i>	Richomme et al. (1991)
Lapachol	Quinoid	<i>L. donovani</i>	Garcia-Granados et al. (1997)
Lauric acid	Lipid	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Licochalcone A	Flavonoid	<i>L. donovani</i>	Cunningham et al. (1972)
		<i>L. donovani</i>	Chen et al. (1993)
		<i>L. donovani</i>	Chen et al. (1994)
		<i>L. donovani</i>	Chen (1995)
Limacine	Alkaloid	<i>L. major</i>	Zhai et al. (1995)
		<i>L. braziliensis</i>	Chan-Bacab and Pena-Rodriguez (2001)
Liriodendronine, N-methyl	Alkaloid	<i>L. donovani</i>	Camacho et al. (2000b)
Liriodenine	Alkaloid	<i>L. amazonensis</i>	Queiroz et al. (1996)
		<i>L. donovani</i>	Waechter et al. (1999)

Table 2. (continued)

Chemical substance ^b	Class	Organism tested	References
Lirioresinol B	Lignan	<i>L. amazonensis</i>	Fevrier et al. (1999)
Loganin, 6-beta hydroxy	Iridoid	<i>L. donovani</i>	Tandon et al. (1991)
Luteolin	Flavonoid	<i>L. donovani</i>	Mittra et al. (2000)
Lysicamine	Alkaloid	<i>L. donovani</i>	Waechter et al. (1999)
Medioresinol, (+)	Lignan	<i>L. amazonensis</i>	Chan-Bacab and Pena-Rodriguez (2001)
Mimengoside A	Triterpene	<i>L. infantum</i>	Emam et al. (1996)
Minquartynoic acid	Lipid	<i>L. major</i>	Rasmussen et al. (2000b)
Molvizarin	Miscellaneous lactone	<i>L. amazonensis</i>	Waechter et al. (1998)
		<i>L. donovani</i>	Sahpaz et al. (1994)
Monomycin	Carbohydrate	<i>L. tropica</i>	Kellina (1964)
		<i>L. tropica</i>	Moskalenko and Pershin (1966)
		<i>L. tropica</i>	Neal (1968)
		<i>L. tropica</i>	Pershin and Moskalenko (1963)
Moschatoline, <i>O</i> -methyl	Alkaloid	<i>L. donovani</i>	Waechter et al. (1999)
Muzanzagenin	Triterpene	<i>L. major</i>	Oketch-Rabah et al. (1997b)
Mycobacillin	Proteid	<i>L. donovani</i>	Chattopadhyay et al. (1976)
Mycophenolic acid	Miscellaneous lactone	<i>L. tropica</i>	Berman and Webster (1982)
Neurolelin B	Sesquiterpene	<i>L. mexicana</i>	Berger et al. (2001)
Neurolelin C	Sesquiterpene	<i>L. mexicana</i>	Berger et al. (2001)
Nyasol, (+)	Lignan	<i>L. major</i>	Oketch-Rabah et al. (1997b)
Nystatin	Miscellaneous lactone	<i>L. donovani</i>	Ghosh (1963a)
		<i>L. donovani</i>	Ghosh (1963b)
		<i>L. donovani</i>	Ghosh and Chaterjee (1961)
		<i>L. donovani</i>	Ghosh and Chaterjee (1963)
Obaberine	Alkaloid	<i>L. braziliensis</i>	Chan-Bacab and Pena-Rodriguez (2001)
Oleanolic acid, epi	Triterpene	<i>L. donovani</i>	Camacho et al. (2000a)
Palmatine	Alkaloid	<i>L. donovani</i>	Vennerstrom et al. (1990)
Palmitic acid	Lipid	<i>L. donovani</i>	Cunningham et al. (1972)
Pannarine	Depside	<i>L. amazonensis</i>	Fournet et al. (1997)
Pannarine, 1'-chloro	Depside	<i>L. amazonensis</i>	Fournet et al. (1997)
Paromomycin	Carbohydrate	<i>L. donovani</i>	Neal and Croft (1984)
		<i>L. tropica</i>	El-On et al. (1985)
		<i>L. tropica</i>	Moskalenko and Pershin (1966)
		<i>L. tropica</i>	Neal (1968)
Paromomycin I	Carbohydrate	<i>L. garnhami</i>	Rezzano de Raffo et al. (1985)
Phaseolinone	Sesquiterpene	<i>L. donovani</i>	Roy et al. (1990)
Phenyl-3-(4-hydroxy-3-(3-methyl-2-butenyl)-phenyl]-prop- <i>trans</i> -2-en-1-one, 1-[2,4-dihydroxy-3-(3-methyl-2-butenyl)]	Flavonoid	<i>L. donovani</i>	Christensen et al. (1994)
Phenylpropane,3,4,5-trimethoxy-8-(3',5'-dimethoxy-4'- <i>trans</i> -propenyl)-phenoxy	Phenylpropanoid	<i>L. donovani</i>	Barata et al. (2000)
Phorbol-13-acetate,12- <i>O</i> -tetradecanol	Diterpene	<i>L. amazonensis</i>	Chan-Bacab and Pena-Rodriguez (2001)

Picroliv	Iridoid	<i>L. donovani</i>	Mittal et al. (1998)
Picrosid I	Iridoid	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Pimaricin	Miscellaneous lactone	<i>L. donovani</i>	Neal and Croft (1984)
Piperine	Alkaloid	<i>L. donovani</i>	Kapil (1993)
Piperogalin	Benzenoid	<i>L. amazonensis</i>	Mahiou et al. (1995)
		<i>L. amazonensis</i>	Fournet et al. (1996)
		<i>L. donovani</i>	Kapil (1993)
Piperone, hydro	Quinoid	<i>L. amazonensis</i>	Mahiou et al. (1996)
Pleiocarpine	Alkaloid	<i>L. donovani</i>	Kam et al. (1999)
Plumbagin	Quinoid	<i>L. amazonensis</i>	Fournet et al. (1992a)
		<i>L. braziliensis</i>	Fournet et al. (1992b)
Plumbagin, 3,3'-bi	Quinoid	<i>L. amazonensis</i>	Fournet et al. (1992a)
		<i>L. braziliensis</i>	Fournet et al. (1992b)
Plumbagin, 8,8'-bi	Quinoid	<i>L. amazonensis</i>	Chan-Bacab and Pena-Rodriguez (2001)
Podocarpusflavone A	Flavonoid	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Podocarpusflavone B	Flavonoid	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Polymyxin B	Proteid	<i>L. tropica</i>	Kellina (1964)
Pregnan-20-one-3-alpha-amino-14-beta-hydroxy	Steroid	<i>L. donovani</i>	Kam et al. (1998)
Puertogaline A	Alkaloid	<i>L. amazonensis</i>	Mahiou et al. (2000)
Puertogaline B	Alkaloid	<i>L. amazonensis</i>	Mahiou et al. (2000)
Pyran, tetrahydro: 2-[beta-para-hydroxy-phenyl-ethyl]-6-(para-hydroxy-phenyl)	Benzenoid	<i>L. amazonensis</i>	Araújo et al. (1998)
Pyrrole-2-carboxaldehyde	Alkaloid	<i>L. mexicana</i>	Compagnone et al. (1999)
Quercetin	Flavonoid	<i>L. donovani</i>	Mittra et al. (2000)
Quinoline, 2-(3,4-dimethoxy-phenyl-ethyl)	Alkaloid	<i>L. amazonensis</i>	Fournet et al. (1993c)
		<i>L. braziliensis</i>	Fournet et al. (1993a)
Quinoline, 2-(3,4-methylenedioxy-phenyl-ethyl)	Alkaloid	<i>L. amazonensis</i>	Fournet et al. (1993c)
		<i>L. braziliensis</i>	Fournet et al. (1994a)
Quinoline, 2- <i>N</i> -pentyl	Alkaloid	<i>L. amazonensis</i>	Fournet et al. (1994a)
		<i>L. braziliensis</i>	Fournet et al. (1993a)
Reserpine	Alkaloid	<i>L. major</i>	Staerk et al. (2000)
Ribenol	Diterpene	<i>L. donovani</i>	Garcia-Granados et al. (1997)
Rigidusine	Diterpene	<i>L. donovani</i>	Morales et al. (2000)
Rollidesin B	Miscellaneous lactone	<i>L. amazonensis</i>	Fevrier et al. (1999)
Rolliniastatin 1	Miscellaneous lactone	<i>L. amazonensis</i>	Carvalho and Ferreira (2001)
Rolliniostatin 2	Miscellaneous lactone	<i>L. amazonensis</i>	Waechter et al. (1998)
		<i>L. donovani</i>	Sahpaz et al. (1994)
Rosenolactone, 6-beta-hydroxy	Diterpene	<i>L. donovani</i>	Chan-Bacab and Pena-Rodriguez (2001)
Rubiadin-1-methyl ether	Quinoid	<i>L. major</i>	Sittie et al. (1999)
Sarachine	Steroid	<i>L. chagasi</i>	Moretii et al. (1998)
		<i>L. major</i>	Sahpaz et al. (1994)
Senegalene	Miscellaneous lactone	<i>L. major</i>	Chan-Bacab and Pena-Rodriguez (2001)
Sepeperine	Alkaloid	<i>L. amazonensis</i>	Mahiou et al. (2000)

Table 2. (continued)

Chemical substance ^b	Class	Organism tested	References
Sergeolide	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Sergeolide, 15-acetyl	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Simalikalactone D	Triterpene	<i>L. donovani</i>	Robert-Gero et al. (1985)
Sinefungin	Alkaloid	<i>L. amazonensis</i>	Haughan et al. (1993)
		<i>L. braziliensis</i>	Avila et al. (1990)
		<i>L. donovani</i>	Lawrence and Robert-Gero (1993)
		<i>L. donovani</i>	Moulay and Robert-Gero (1995)
		<i>L. donovani</i>	Phelouzat et al. (1995)
		<i>L. donovani</i>	Neal et al. (1989)
		<i>L. mexicana</i>	Haughan et al. (1993)
		<i>L. tropica</i>	Paolantonacci et al. (1987)
Skimmianine	Alkaloid	<i>L. amazonensis</i>	Fournet et al. (1993c)
		<i>L. braziliensis</i>	Fournet et al. (1993a)
		<i>L. donovani</i>	Fournet et al. (1994a)
Squamocin	Miscellaneous lactone	<i>L. amazonensis</i>	Waechter et al. (1998)
		<i>L. major</i>	Sahpaz et al. (1994)
Striatin A	Diterpene	<i>L. amazonensis</i>	Inchausti et al. (1997)
Striatin B	Diterpene	<i>L. amazonensis</i>	Inchausti et al. (1997)
Sulfuretin	Flavonoid	<i>L. donovani</i>	Kayser et al. (1999)
Surinamensine	Lignan	<i>L. donovani</i>	Barata et al. (2000)
Sylvaticin	Miscellaneous lactone	<i>L. amazonensis</i>	Fevrier et al. (1999)
Taurin, 8- α -hydroxy	Sesquiterpene	<i>L. donovani</i>	Cubukcu et al. (1998)
Taxol	Diterpene	<i>L. major</i>	Doherty et al. (1998)
Tetradeca-7-11-dienoic acid, 3-6-epidioxy-4,6,8,10-tetraethyl	Lipid	<i>L. mexicana</i>	Compagnone et al. (1998)
Tetrandrine, iso	Alkaloid	<i>L. amazonensis</i>	Fournet et al. (1993d)
Titucall-3- α -hydroxy-7,24-Z-dien-26-oic acid	Triterpene	<i>L. donovani</i>	Camacho et al. (2000a)
Tyrocidine	Proteid	<i>L. tropica</i>	Weinman (1943)
Unonopsine	Alkaloid	<i>L. donovani</i>	Waechter et al. (1999)
Ursolic acid	Triterpene	<i>L. amazonensis</i>	Sauvain et al. (1993)
Ushinsunine, nor	Alkaloid	<i>L. amazonensis</i>	Queiroz et al. (1996)
Usnic acid	Oxygen heterocycle	<i>L. amazonensis</i>	Fournet et al. (1997)
Verbascoside	Phenylpropanoid	<i>L. infantum</i>	Emam et al. (1995)
Verbascoside, iso	Phenylpropanoid	<i>L. infantum</i>	Emam et al. (1995)
Vernodalol	Sesquiterpene	<i>L. infantum</i>	Koshimizu et al. (1994)
Vernodalol	Sesquiterpene	<i>L. infantum</i>	Koshimizu et al. (1994)
Vernolide	Sesquiterpene	<i>L. infantum</i>	Koshimizu et al. (1994)
Vernolide, hydroxy	Sesquiterpene	<i>L. infantum</i>	Koshimizu et al. (1994)
Voacangine	Alkaloid	<i>Leishmania</i> sp.	Bou Habib et al. (1998)
With-5-enolide D, 18-acetoxy-5,6-deoxy	Steroid	<i>L. brasiliensis</i>	Bravo et al. (2001)
Yohimbine, α	Alkaloid	<i>L. major</i>	Staerk et al. (2000)
Zaluzanin C, dehydro	Sesquiterpene	<i>L. amazonensis</i>	Fournet et al. (1993b)

^aOnly positive chemical compounds tested for antileishmanial activity were presented in Table 2. Data for negative results is available from the authors on request.

^bIn most cases the reference compound to a positive control were either glucantime or pentamidine.

arbortristis) showed more than 75% inhibition (at 1 g/kg/day \times 5 orally) of multiplication of parasites on day 7 and/or 28 post treatment with an increased survival period (Singha et al., 1992).

Extracts of 11 plants used in Nigerian traditional medicine have been evaluated for possible antileishmanial activity using a radiorespirometric microtest technique based on *in vitro* inhibition of catabolism of $^{14}\text{CO}_2$ from a battery of ^{14}C -substrates by promastigotes. Of 13 methanol extracts tested, 5 from *Gongronema latifolia*, *Dorstenia multiradiata*, *Picralima nitida*, *Cola attiensis*, and *Desmodium gangeticum* were active at concentrations of 50 $\mu\text{g}/\text{ml}$ or less against a visceral *Leishmania* isolate (Iwu et al., 1992).

Crude extracts collected from different parts of Spain (60 representing 12 species from 7 families) have been screened for antiparasitic activity against *L. infantum*, and 30% of the extracts showed activity. The most promising extracts originate from plants used in traditional medicine, such as *Inula montana*, *Bupleurum rigidum* and *Scrophularia scorodonia* (Martin et al., 1998).

The drugs used currently for treatment of Kala-azar, sodium stibogluconate (SSG) and pentamidine cause severe toxic side-effects and acute immunosuppression in the treated individuals. Picroliv, a standardized mixture of iridoid glycosides, prepared from the alcoholic extract of the root and rhizome of *Picrorhiza kurroa*, has shown strong hepatoprotective activity against several models of hepatotoxicity. Therefore, this study was undertaken to study the effects of picroliv (12.5 mg/kg body wt. \times 7 days oral) alone and in combination with SSG on parasitemia, lipid peroxidation and hepatic marker enzymes of golden hamsters during *L. donovani* infection. The results indicated a marked hepatoprotective effect of picroliv in terms of biochemical markers, and a significant antileishmanial activity, implying that it can be utilized as an adjuvant to chemotherapy or in combination therapy of Kala-azar along with SSG, thus enhancing the efficacy of antileishmanials (Mittal et al., 1998).

The ethanolic extract of *Yucca filamentosa*, showed potent activity against *L. amazonensis* at a concentration of 5 $\mu\text{g}/\text{ml}$ (Plock et al., 2001). Other plants with marked activity against *L. donovani* were *Khaya senegalensis* and *Anthostema senegalense* with IC_{50} values of 9.8 and 9.1 $\mu\text{g}/\text{ml}$, respectively (Abreu et al., 1999).

Studies carried out in Colombia with *Annona muricata* against *L. braziliensis* and *L. panamenis* showed that its activity was greater than that of meglumine antimoniate (Glucantime[®]) (Jaramillo et al., 2000). These results show the importance of the investigation of plants with therapeutic potential in the treatment of Leishmaniases.

We found 101 plants described in the literature with antileishmanial activity. The plants are listed in Table 1

in alphabetical order of their family, scientific names, country, plant part used, kind of extract, result and references.

Antileishmanial activity of chemically defined molecules

Four bisbenzisoquinoline alkaloids, antioquine, berbamine, gyrocarpine and isotetrandrine, were tested in BALB/c mice infected with *L. amazonensis*. The treatments were initiated 1 day after the parasitic infection, with alkaloid at 100 mg/kg body wt./body wt./day for 14 days and the reference compound, Glucantime[®] at 200 mg/kg body wt./body wt./day. Antioquine, berbamine and gyrocarpine were less potent than Glucantime against *L. amazonensis*. Only isotetrandrine exhibited activity approximately equal to or greater than Glucantime (Fournet et al., 1993d).

A series of aurones with drug-potential for *Leishmania* sp. infections was identified *in vitro* using both a direct cytotoxicity assay against extracellular promastigotes of *L. donovani*, *L. infantum*, *L. enriettii*, and *L. major*, and a test against intracellular amastigote forms of *L. donovani* residing within murine macrophages. The most active aurone, 6-hydroxy-2-[phenylmethylene]-3(2H)-benzofuranone had an IC_{50} of 0.45 $\mu\text{g}/\text{ml}$ in the extra-, and IC_{50} value of 1.40 $\mu\text{g}/\text{ml}$ in the intracellular assay. Other aurones were active between 0.06–12.50 $\mu\text{g}/\text{ml}$ and 0.04–7.81 $\mu\text{g}/\text{ml}$, respectively. When tested against murine bone marrow-derived macrophages as a mammalian host cell control, the compounds showed only moderate cytotoxicity (IC_{50} value 2.32 to <25.0 $\mu\text{g}/\text{ml}$) (Kayser and Kiderlen, 1999).

From the hexane extract of roots of *Annona haematantha* an α,β -unsaturated δ -lactone was isolated and identified as argentilactone. This compound exhibited *in vitro* activity against various strains of *Leishmania* sp. at 10 $\mu\text{g}/\text{ml}$. BALB/c mice infected with *L. amazonensis* were treated 4 weeks after infection with argentilactone by oral or subcutaneous routes for 14 days at 25 mg/kg body wt. daily. The reference drug, *N*-methylglucamine antimonate, was administered by subcutaneous injections at 100 mg/kg body wt. for 14 days. Under these conditions, argentilactone showed the same efficacy as the reference drug, reducing by 96% the parasite loads in the lesion and by 50% the parasite burden in the spleen (Waechter et al., 1997).

Compounds isolated from plants with promising activity against the *Leishmania* genera and low toxicity as compared to the pentavalent antimonial drugs include chimanine B; 4-hydroxy-tetralone; 8-8'-biplyumbagin; rolliniastatin-1; squamocin; dictyolamide A and B and 2-benzoxazolinone (Carvalho and Ferreira, 2001).

Reviews by Carvalho and Ferreira (2001), Chan-Bacab and Pena-Rodriguez (2001) provide further

information on the antileishmanial activity of chemically defined molecules.

We encountered 239 chemically defined natural molecules reported in the literature, which have been evaluated for antileishmanial activity. The active compounds, which have been isolated and identified, belong to the classes of alkaloids (68), triterpenes (29), sesquiterpenes (19), miscellaneous lactones (18), quinoids (16), flavonoids (13), diterpenes (13), steroids (10), lipids (8), iridoids (8), oxygen heterocycles (7), benzenoids (6), carbohydrates (5), lignans (5), proteids (4), coumarins (3), phenylpropanoids (3), depsides (2), a sulfur compound (1), and a monoterpene (1). The compounds are arranged in Table 2 in alphabetical order. Each entry gives the following information in sequence: chemical name, class, organism tested, and references.

Conclusion

The present study shows a range of plant extracts exhibit interesting antileishmanial properties *in vitro*, seeming to validate their use in folk medicine. The potent leishmanicidal activities of certain chemically defined molecules isolated from natural origins represent an exciting advance in the search for novel antiprotozoal agents at a time when there is an urgent need for new innovative drug leads.

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References

- Abreu, P.M., Martins, E.S., Kayser, O., Bindseil, K.U., Siems, K., Seemann, A., Frevet, J., 1999. Antimicrobial, antitumor and antileishmanial screening of medicinal plants from Guinea-Bissau. *Phytomedicine* 6, 187–195.
- Araújo, C.A.C., Alegrio, L.V., Leon, L.L., 1998. Antileishmanial activity of compounds extracted and characterized from *Centrolobium sclerophyllum*. *Phytochemistry* 49, 751–754.
- Ashford, R.W., 2000. The leishmaniasis as emerging and reemerging zoonoses. *Int. J. Parasitol.* 30, 1269–1281.
- Avila, J.L., Rojas, T., Monzon, H., Convit, J., 1990. Sinefungin as treatment for American Leishmania in sensitive BALB/c and resistant c57BL/6 mice. *Am. J. Trop. Med. Hyg.* 43, 139–145.
- Balana, F.R., Reguera, R., Cubria, J.C., Ordonez, D., 1998. The pharmacology of leishmaniasis. *Gen. Pharmacol.* 30, 435–443.
- Barata, L.E.S., Santos, L.S., Ferri, P.H., Phillipson, F.J.D., Paine, A., Croft, S.L., 2000. Antil-leishmanial activity of neoligands from *Virola* species and synthetic analogues. *Phytochemistry* 55, 589–595.
- Barral-Neto, M., Von Sohsten, R.L., Teixeira, M., Conrado dos Santos, W.L., Pompeu, M.L., Moreira, R.A., Oliveira, J.T.A., Cavada, B.S., Falcoff, E., Barral, A., 1996. *In vivo* protective effect of the lectin from *Canavalia brasiliensis* of BALB/c mice infected by *Leishmania amazonensis*. *Acta Trop.* 60, 237–250.
- Berger, I., Passreiter, C.M., Cáceres, A., Kubelka, W., 2001. Antiprotozoal activity of *Neurolaena lobata*. *Phyther. Res.* 15, 327–330.
- Berman, J.D., Webster, H.K., 1982. *In vitro* effects of mycophenolic acid and allopurinol against *Leishmania tropica* in human macrophages. *Antimicrob. Agents Chemother.* 21, 887–891.
- Bhadra, R., 1993. Antileishmanial agents. *Drugs Future* 18, 451–463.
- Bhakuni, D.S., Goel, A.K., Jain, S., Mehrotra, B.N., Patnaik, G.K., Prakash, V., 1988. Screening of Indian plants for biological activity. Part XIII. *Indian J. Exp. Biol.* 26, 883–904.
- Bodley, A.L., Shapiro, T.A., 1995. Molecular and cytotoxic effects of camptothecin, a topoisomerase I inhibitor on *Trypanosomes* and *Leishmania*. *Proc. Natl. Acad. Sci. (USA)* 92, 3726–3730.
- Bodley, A.L., Mc Garry, M.W., Shapiro, T.A., 1995. Drug cytotoxicity assay for African *Trypanosomes* and *Leishmania* species. *J. Infect. Dis.* 172, 1157–1159.
- Bou Habib, D.C., Ferraro, G.A., Bou Habib, E.M.S., Delorenzi, J.C.M.O.B., Pinto, A.D.C., Moraes de Rezende, C., Teixeira de Andrade, M., 1998. Alkaloids of *Peschiera australis* for use in treatment of leishmaniasis. *Patent Brazil* 98 04,032, 49.
- Bravo, J.A., Sauvain, M., Gimenez, A., Munoz, V., Callapa, J., Le Men-Olivier, L., Massiot, G., Lavaud, C., 1999. Bioactive phenolic glycosides from *Amburana cearensis*. *Phytochemistry* 50, 71–74.
- Bravo, J.A., Sauvain, M., Gimenez, T.A., Balanza, E., Serani, L., Laprevote, O., Massiot, G., Lavaud, C., 2001. Typanocidal withanolides and withnolide glycosides from *Dunalia brachyacantha*. *J. Nat. Prod.* 64, 720–725.
- Bringman, G., Hamm, A., Gunther, C., Michel, M., Brun, R., Mudogo, V., 2000. Ancistrocalin A and B, two new bioactive naphthlisoquinolines, and related naphthoic acids from *Ancistrocladus ealaensis*. *J. Nat. Prod.* 63, 1465–1470.
- Camacho, M.D.R., Mata, R., Castaneda, P., Kirby, G.C., Warhust, D.C., Croft, S.L., Phillipson, J.D., 2000a. Bioactive compounds from *Celaenodredon mexicanum*. *Planta Med.* 66, 462–468.
- Camacho, M.D.R., Kirby, G.C., Warhust, D.C., Croft, S.L., Phillipson, J.D., 2000b. Oxoaporphine alkaloids and quinines from *Stephanina dinklagei* and evaluation of their antiprotozoal activities. *Planta Med.* 66, 478–480.

- Carson, D.A., Chang, K.P., 1981. Phosphorylation and antileishmanial activity of formycin B. *Biochem. Biophys. Res. Commun.* 100, 1377–1383.
- Carvalho, P.B., Ferreira, E.I., 2001. Leishmaniasis phytotherapy. Nature's leadership against an ancient disease—review. *Fitoterapia* 72, 599–618.
- Carvalho, P.B., Arribas, M.A.G., Ferreira, E.I., 2000. Leishmaniasis. What do we know about its chemotherapy? *Rev. Bras. Ci. Farm* 36 (Suppl. 1), 69–96.
- Cavé, A., Hocquemiller, R., Laprovote, O., 1990. Isolation of acetogenins and their use as antiparasitic agents. *Patent-Fr Demande-2,634,123*
- Chalcat, P., Colas Belcour, J., Destombes, P., Drouhet, E., Fromentin, H., Martin, L., Ravisse, P., Silverie, J., 1965. A case of mucus skin leishmaniasis resistant to antimony compounds and cleared by amphotericin B. *Bull. Soc. Pathol. Exot.* 58, 73–80.
- Chan-Bacab, M.J., Pena-Rodriguez, L.M., 2001. Plant natural products with leishmanicidal activity. *Nat. Prod. Rep.* 18, 674–688.
- Chance, M.L., 1995. New developments in the chemotherapy of leishmaniasis. *Ann. Trop. Med. Parasitol.* 89, 37–43.
- Chattopadhyay, T.K., Ghatak, S.N., Bhaduri, N.A., Ghosh, D.K., 1976. Antileishmanial activity of mycobacillin. *J. Antibiot.* 30, 262–266.
- Chen, M., 1995. Antileishmanial and antimalarial activities of licochalcone A. *Dan. Med. Bull.* 42, 378–380.
- Chen, M., Christensen, S.B., Blom, J., Lemmich, E., Nadelmann, L., Fich, K., Theander, T.G., Kharazmi, A., 1993. Licochalcone A, a novel antiparasitic agent with potent activity against human pathogenic protozoan species of *Leishmania*. *Antimicrob. Agents Chemother.* 37, 2550–2556.
- Chen, M., Christensen, S.B., Theander, T.G., Kharazmi, A., 1994. Antileishmanial activity of licochalcone A in mice infected with *Leishmania major* and in hamsters infected with *Leishmania donovani*. *Antimicrob. Agents Chemother.* 38, 1339–1344.
- Christensen, S.B., Ming, C., Andersen, L., Hjerne, U., Olsen, C.E., Cornett, C., Theander, T.G., Kharazmi, A., 1994. An antileishmanial chalcone from Chinese licorice roots. *Planta Med.* 60, 121–123.
- Compagnone, R.S., Pina, I.C., Rangel, H.R., Dagger, F., Suarez, A.I., Reddy, M.V.R., Faulkner, D.J., 1998. Antileishmanial cyclic peroxides from the Palauan sponge *Plakortis aff. angulospiculatus*. *Tetrahedron* 54, 3057–3068.
- Compagnone, R.S., Oliveri, M.C., Pina, I.C., Marques, S., Rangel, H.R., Dager, F., Suarez, A.L., Gomez, M., 1999. 5-Alkylpyrrole-2-carboxaldehydes from the Caribbean sponges *Mycale microsigmatosa* and *Desmapsamma anchorata*. *Nat. Prod. Lett.* 13, 203–211.
- Cook, J.A., Holbrook, T.W., Parker, B.W., 1980. Visceral leishmaniasis in mice: protective effect of glucan. *J. Reticuloendothel. Soc.* 27, 567–573.
- Cubukcu, B., Gasquet, M., Delmas, F., Favel, A., Mericli, A.H., Balansard, G., 1998. Evaluation of antiprotozoal and antigungal activities of Turkish *Artemisia santonicum*. *J. Fac. Pharm. Istanbul Univ.* 32, 20–22.
- Cunningham, L.V., Kazan, B.H., Kuwahara, S.S., 1972. Effect of long-chain fatty acids on some trypanosomatid flagellates. *J. Gen. Microbiol.* 70, 491–496.
- Da Silva, S.A.G., Costa, S.S., Mendonça, S.C.F., Silva, E.M., Moraes, V.L.G., Rossi-Bergmann, B., 1995. Therapeutic effect of oral *Kalanchoe pinnata* leaf extract in murine leishmaniasis. *Acta Trop.* 60, 201–210.
- Delorenzi, J.C., Attias, M., Gattass, C.R., Abdrade, M., Rezende, C., Cunha-Pinto, A., Henrique, A.T., Bou-Habib, D.C., Saraiva, E.M., 2001. Antileishmanial activity of an indole alkaloid from *Peschiera australis*. *Antimicrob. Agents Chemother.* 45, 1349–1354.
- Demirci, F., Demirci, B., Ali, S.A., Shoudary, M.I., Baser, K.H.C., 1998. Bioassays on *Periploca graeca* L. (Silk Vine). *Acta Pharm. Turc* 40, 145–149.
- Doherty, T.M., Sher, A., Vogel, S.N., 1998. Paclitaxel (Taxol)-induced killing of *Leishmania major* in murine macrophages. *Infect Immun.* 66, 4553–4556.
- Edward, J.A.M., Amador, A.T., Luis, G.P., Stell, T.D.Y., Santiago, N., 1995. *In vivo* activity of berberine chloride towards cutaneous leishmaniasis caused by *Leishmania mexicana* 856/INS. *Rev. Colomb. Quim.* 24, 55–64.
- El-On, J., Weinrauch, L., Livshin, R., Even-Paz, Z., Jacobs, G.P., 1985. Topical treatment of recurrent cutaneous leishmaniasis with ointment containing paromomycin and methylbenzethonium chloride. *Br. Med. J.* 291, 704–705.
- Emam, A.M., Moussa, A.M., Essa, M.A., Faure, R., Elias, R., Balansard, G., Boudon, G., 1995. Isolation of phenyl propanoid glycosides from *Buddleja madagascariensis* Lam. leaves. *Pharm. Pharmacol. Lett.* 5, 177–178.
- Emam, A.M., Moussa, A.M., Faure, R., Favel, A., Delmas, F., Elias, R., Balansard, G., 1996. Isolation and biological study of a triterpenoid saponin, mimengoside A, from the leaves of *Buddleja madagascariensis*. *Planta Med.* 62, 92–93.
- Fevrier, A., Ferreira, M.E., Fournet, A., Yaluff, G., Inchausti, A., Arias, A.R., Hocquemiller, R., Waechter, A.I., 1999. Acetogenins and other compounds from *Rollinia emarginata* and their antiprotozoal activities. *Planta Med.* 65, 47–49.
- Fournet, A., Barrios, A.A., Munoz, V., Hocquemiller, R., Cavé, A., 1992a. Effect of natural naphthoquinones in BALB/c mice infected with *Leishmania amazonensis* and *L. venezuelensis*. *Trop. Med. Parasitol.* 43, 219–222.
- Fournet, A., Angelo, A., Munoz, V., Roblot, F., Hocquemiller, R., Cavé, A., 1992b. Biological and chemical studies of *Pera benensis*, a Bolivian plant used in folk medicine as treatment of cutaneous leishmaniasis. *J. Ethnopharmacol.* 37, 159–164.
- Fournet, A., Hocquemiller, R., Roblot, F., Cavé, A., Richomme, P., Bruneton, J., 1993a. Chimanines, novel substituted quinoleines, isolated from Bolivian antiparasitic plant, *Galipea longiflora*. *J. Nat. Prod.* 56, 1547–1552.
- Fournet, A., Munoz, V., Roblot, F., Hocquemiller, R., Cavé, A., Gantier, J.C., 1993b. Antiprotozoal activity of dehydrozaluzanin C, a sesquiterpene lactone isolated from *Munnozia maronii* (Asteraceae). *Phytother. Res.* 7, 111–115.
- Fournet, A., Barrios, A.A., Munoz, V., Hocquemiller, R., Cavé, A., Bruneton, J., 1993c. 2-Substituted quinoline alkaloids as potential antileishmanial drugs. *Antimicrob. Agents Chemother.* 37, 859–863.
- Fournet, A., Barrios, A.A., Munoz, V., Hocquemiller, R., Cavé, A., 1993d. Effect of some bisbenzylisoquinoline

- alkaloid on American *Leishmania* sp. in BALB/c mice. *Phytother. Res.* 7, 281–284.
- Fournet, A., Barrios, A.A., Munoz, V., Hocquemiller, R., Roblot, F., Cavé, A., Richomme, P., Bruneton, J., 1994a. Antiprotozoal activity of quinoline alkaloids isolated from *Galipea longiflora*, a Bolivian plant used as a treatment for cutaneous leishmaniasis. *Phytother. Res.* 8, 174–178.
- Fournet, A., Barrios, A.A., Munoz, V., 1994b. Leishmanicidal and trypanocidal activities of Bolivian medicinal plants. *J. Ethnopharmacol.* 41, 19–37.
- Fournet, A., Ferreira, M.E., Arias, A.R., Fuentes, S., Torres, S., Inchausti, A., Yaluff, A., Nakayama, H., Mahiou, V., Hocquemiller, R., Cavé, A., 1996. *In vitro* and *in vivo* leishmanicidal studies of *Peperomia galioides* (Piperaceae). *Phytomedicine* 3, 271–275.
- Fournet, A., Ferreira, M.E., Arias, A.R., Ortiz, S.T., Inchausti, A., Yaluff, G., Quilhot, W., Fernandez, E., Hidalgo, M.E., 1997. Activity of compounds isolated from Chilean lichens against experimental cutaneous leishmaniasis. *Comp. Biochem. Physiol. Ser. C* 116, 51–54.
- Fuchino, H., Koide, T., Takahashi, M., Sekita, S., Satake, M., 2001. New sesquiterpene lactones from *Elephantopus mollis* and their leishmanicidal activities. *Planta Med.* 67, 647–653.
- Furtado, T.A., Cisalpino, E.O., Santos, U.M., 1960. *In vitro* studies of effect of amphotericin B on *Leishmania brasiliensis*. *Antibiot. Chemother.* 10, 692–693.
- Garcia-Granados, A., Linan, E., Martinez, A., Rivas, F., Mesa-Valle, C., Castilla-Calvente, J.J., Osuna, A., 1997. *In vitro* action of ent-manoyl oxides against *Leishmania donovani*. *J. Nat. Prod.* 60, 13–16.
- Ghazanfari, T., Hassan, Z.M., Ebtekar, M., Ahmadiani, A., Naderi, G., Azar, A., 2000. Garlic induces a shift in cytokine in *Leishmania major*—infected balb/mice. *Scand. J. Immunol.* 52, 491–495.
- Ghosh, B.K., 1963a. Action of an antifungal antibiotic, nystatin, on the protozoa *Leishmania donovani*. Studies on the cytological and cytochemical changes. *Ann. Biochem. Exp. Med.* 23, 193–200.
- Ghosh, B.K., 1963b. Action of an antifungal antibiotic, nystatin, on the protozoa, *Leishmania donovani*. Investigations concerning the effect on isolated member and the isolation of antibiotic-rich cell portions of *L. donovani*. *Ann. Biochem. Exp. Med.* 23, 337–344.
- Ghosh, B.K., Chatterjee, A.N., 1961. Action of an antifungal antibiotic, nystatin, on the protozoa *Leishmania donovani*. Studies on the metabolism of *Leishmania donovani*. *Ann. Biochem. Exp. Med.* 21, 307–322.
- Ghosh, B.K., Chatterjee, A.N., 1963. Action of an antifungal antibiotic, nystatin, on the protozoa *Leishmania donovani*. Studies on the lysis of the cells of *Leishmania donovani*. *Ann. Biochem. Exp. Med.* 23, 173–186.
- Guerrero, C., Taboada, J., Biaz, J.B., Oliva, A., Ortega, A., 1995. Incomptins A and B, two heliangolides from *Decachaeta incompta*. Preliminary studies on the biological activity of incomptin B. *Rev. Latinoam. Quim.* 23, 142–147.
- Hatimi, S., Boudouma, M., Bichichi, M., Chaib, N., Idrissi, N.G., 2001. *In vitro* evaluation of antileishmania activity of *Artemisia herba alba* Asso. *Bull. Soc. Pathol. Exot.* 94, 29–31.
- Haughan, P.A., Chance, M.L., Goad, L.J., 1993. Effects of simefungin on growth and sterol composition of *Leishmania* promastigotes. *Exp. Parasitol.* 77, 147–154.
- Hazra, B., Ghosh, R., Banerjee, A., Kirby, G.C., Warhurst, D.C., Phillipson, J.D., 1995. *In vitro* antiplasmodial effects of diospyrin, a plant-derived naphthoquinoid, and a novel series of derivatives. *Phytother. Res.* 9, 72–74.
- Hiraoka, O., Satake, H., Iguchi, S., Matsuda, A., Ueda, T., Wataya, Y., 1986. Carbocyclic inosine as a potent antileishmanial agent: the metabolism and selective cytotoxic effects of carbocyclic inosine in promastigotes of *Leishmania tropica* and *Leishmania donovani*. *Biochem. Biophys. Res. Commun.* 134, 1114–1121.
- Hocquemiller, R., Cortex, D., Arango, G.J., Myint, S.H., Cavé, A., Angel, A., Munos, V., Fournet, A., 1991. Isolation and synthesis of espintanol, a novel antiparasitic monoterpene. *J. Nat. Prod.* 54, 443–452.
- Inchausti, A., Yaluff, G., Arias, A.R., Torres, S., Ferreira, M.E., Nakayama, H., Schinini, A., Lorenzen, K., Anke, T., Fournet, A., 1997. Leishmanicidal and trypanocidal activity of extracts and secondary metabolites from Basidiomycetes. *Phytother. Res.* 11, 193–197.
- Iwu, M.M., Jackson, J.E., Tally, J.D., Klayman, D.L., 1992. Evaluation of plant extracts for antileishmanial activity using a mechanism-based radiorespirometric microtechnique (RAM). *Planta Med.* 58, 436–441.
- Jaramillo, M.C., Arango, G.J., González, M.C., Robledo, S.M., Velez, I.D., 2000. Cytotoxicity and antileishmanial activity of *Annona muricata* pericarp. *Fitoterapia* 71, 183–186.
- Kam, T.S., Sim, K.M., Koyano, T., Toyoshima, M., Hayashi, M., Komyiama, K., 1998. Cytotoxic and leishmanicidal aminoglycosideroids and aminosteroids from *Holarrhena curtisii*. *J. Nat. Prod.* 61, 1332–1336.
- Kam, T.S., Sim, K.M., Koyano, T., Komyiama, K., 1999. Leishmanicidal alkaloids from *Kopsia griffithii*. *Phytochemistry* 50, 75–79.
- Kapil, A., 1993. Piperine: a potent inhibitor of *Leishmania donovani* promastigotes *in vitro*. *Planta Med.* 59, 474.
- Kayser, O., Kiderlen, A.F., 1999. Leishmanicidal activity of auronones. *Tokai J. Exp. Clin. Med.* 23, 423–426.
- Kayser, O., Kiderlen, A.F., 2001. *In vitro* leishmanicidal activity of naturally occurring chalcones. *Phytother. Res.* 15, 148–152.
- Kayser, O., Kiderlen, A.F., Folkens, U., Koldziej, H., 1999. *In vitro* leishmanicidal activity of auronones. *Planta Med.* 65, 316–319.
- Kayser, O., Kolodziej, H., Kiderlen, A.F., 2001a. Immunomodulatory principles of *Pelargonium sidoides*. *Phytother. Res.* 15, 122–126.
- Kayser, O., Kiderlen, A.F., Bertels, S., Siems, K., 2001b. Antileishmanial activities of aphidicolin and its semisynthetic derivatives. *Antimicrob. Agents Chemother.* 45, 288–292.
- Kellina, O.I., 1964. Chemotherapeutic activity of certain preparations in experimental cutaneous leishmaniasis of white mice. *Probl. Med. Parazitol. I Profilaktiki Infekcii Moscow*, 317–338.

- Khalid, S.A., Friedrichsen, G.M., Kharazmi, A., Theander, T.G., Olsen, C.E., Christensen, S.B., 1998. Limonoids of *Khaya senegalensis*. *Phytochemistry* 49, 1769–1772.
- Koshimizu, K., Ohigashi, H., Huffman, M.A., 1994. Use of *Vernonia amygdalina* by wild chimpanzee: possible roles of its bitter and related constituents. *Physiol. Behav.* 56, 1209–1216.
- Lavaud, C., Massiot, G., Barrera, J.B., Moret, C., Le Men-Olivier, L., 1994. Triterpene saponins from *Myrsine pellucida*. *Phytochemistry* 37, 1671–1677.
- Lavaud, C., Massiot, G., Vasquez, C., Moret, C., Sauvain, M., Balderrama, L., 1995. 4-Quinolinone alkaloids from *Dictyoloma peruviana*. *Phytochemistry* 40, 317–320.
- Lawrence, F., Robert-Gero, M., 1993. Distribution of macromolecular methylations in promastigotes of *Leishmania donovani* and impact of sinefungin. *J. Eukaryot. Microbiol.* 40, 581–589.
- Lemma, A., Maxwell, A., Brody, G., 1972. Antimicrobial and anthelmintic activities of endod (*Phytolacca dodecandra*). Research Report No. 12, Inst. Pathobiol., Addis Ababa Univ., pp. 6–10.
- Lockman, Y., Vardy, D., Ohayon, D., El-On, J., 1991. The failure of traditionally used desert plants to act against cutaneous leishmaniasis in experimental animals. *Ann. Trop. Med. Parasitol.* 85, 499–501.
- Lyubimova, L.K., Fateeva, L.I., Sergeeva, L.A., 1979. Experimental characteristics of the antileishmaniasis action of amphotericin B. *Antibiotiki (Moscow)* 24, 281–284.
- Mahiou, V., Roblot, F., Hocquemiller, R., Cavé, A., Arias, R., Inchausti, A., Yaluff, G., Fournet, A., Angelo, A., 1994. New aporphine alkaloids from *Guatteria foliosa*. *J. Nat. Prod.* 57, 890–895.
- Mahiou, V., Roblot, F., Hocquemiller, R., Cavé, A., Barrios, A.A., Fournet, A., Ducrot, P.H., 1995. Piperogalin, a new prenylated diphenol from *Peperomia galioides*. *J. Nat. Prod.* 58, 324–328.
- Mahiou, V., Roblot, F., Hocquemiller, R., Cavé, A., Arias, A.R., Inchausti, A., Yaluff, G., Fournet, A., 1996. New prenylated quinones from *Peperomia galioides*. *J. Nat. Prod.* 59, 694–697.
- Mahiou, V., Roblot, F., Fournet, A., Hocquemiller, R., 2000. Bisbenzylisoquinoline alkaloids from *Guatteria boliviana* (Annonaceae). *Phytochemistry* 54, 709–716.
- Martin, T., Villaescusa, L., Gasquet, M., Delma, F., Bartolome, C., Diaz-Lanza, A.M., Ollivier, E., Balansard, G., 1998. Screening for protozoocidal activity of Spanish plants. *Pharm. Biol.* 36, 56–62.
- McClure, C.D., Nolan, L.L., Zatyryka, S.A., 1996. Antileishmanial properties of *Allium sativum* extracts and derivatives. *Acta Hort.* 426, 183–191.
- Medda, S., Mukhopadhyay, S., Basu, M.K., 1999. Evaluation of the *in vivo* activity and toxicity of amarogentin, an antileishmanial agent, in both liposomal and niosomal forms. *J. Antimicrob. Chemother.* 44, 791–794.
- Mittal, N., Gupta, N., Saksena, S., Goyal, N., Roy, U., Rastogi, A.K., 1998. Protective effect of picroliv from *Picrorhiza kurroa* against *Leishmania donovani* infections in *Mesocricetus auratus*. *Life Sci.* 63, 1823–1834.
- Mittra, B., Saha, A., Chowdhury, A.R., Pal, C., Mandal, S., Mukhopadhyay, S., Bandyopadhyay, S., Majumder, H.K., 2000. Luteolin, an abundant dietary component is a potent anti-leishmanial agent that acts by inducing topoisomerase II-mediated kinetoplast DNA cleavage leading to apoptosis. *Mol. Med. New York* 6, 527–541.
- Moideen, S.V.K., Houghton, P.J., Croft, S.L., 1997. Antileishmanial activity of *Kigelia pinnata* root bark constituents. *J. Pharm. Pharmacol.* 49, 114.
- Morales, G., Sierra, P., Borquez, J., Loyola, L.A., 2000. Rigidusine, a new glycoditerpenoid from *Haplopappus rigidus*. *Bol. Soc. Chil. Quim.* 45, 611–614.
- Moret, C., Sauvain, M., Lavaud, C., Massiot, G., Bravo, J.A., Munoz, V., 1998. A novel antiprotozoal aminosteroid from *Saracha punctata*. *J. Nat. Prod.* 61, 1390–1393.
- Moskalenko, N.Y., Pershin, G.N., 1966. Comparative chemotherapeutic effects of paromomycin and monomycin in induced leishmaniasis of mouse skin. *Farmakol. Toksikol.* 29, 90–94.
- Moulay, L., Robert-Gero, M., 1995. *Leishmania donovani*: enhanced expression of soluble acid phosphatase in the presence of sinefungin, an antiparasitic agent. *Exp. Parasitol.* 80, 8–14.
- Mshvildadze, V., Favela, A., Delmas, F., Elias, R., Faure, R., Decanosidze, G., Kemertelidze, E., Balansard, G., 2000. Antifungal and antiprotozoal activities of saponins from *Hedera colchica*. *Pharmazie* 55, 325–326.
- Mukherjee, T., Roy, K., Bhaduri, A., 1990. Acivicin: a highly active potential chemotherapeutic agent against visceral leishmaniasis. *Biochem. Biophys. Res. Commun.* 170, 426–432.
- Munos, M.H., Mayrargue, J., Fournet, A., Gantier, J.C., Hocquemiller, R., Moskowitz, H., 1994. Synthesis of an antileishmanial alkaloid isolated from *Galipea longiflora*, a Bolivian plant used as treatment for cutaneous leishmaniasis. *Phytother. Res.* 8, 174–178.
- Munoz, V., Moret, C., Sauvain, M., Caron, C., Prozel, A., Massiot, G., Richard, B., Le Men-Olivier, L., 1994. Isolation of bis-indole alkaloids with antileishmanial and antibacterial activities from *Peschiera* var *heurkii* (Syn. *Tabernaemontana* var *heurkii*). *Planta Med.* 60, 455–459.
- Neal, R.A., 1968. The effect of antibiotics of the neomycin group on experimental cutaneous leishmaniasis. *Ann. Trop. Med. Parasitol.* 62, 54–62.
- Neal, R.A., 1970. Effect of emetine and related compounds on experimental cutaneous leishmaniasis. *Ann. Trop. Med. Parasitol.* 64, 159–165.
- Neal, R.A., Croft, S.L., 1984. An *in vitro* system for determining the activity of compounds against the intracellular amastigote form of *Leishmania donovani*. *J. Antimicrob. Chemother.* 14, 463–475.
- Neal, R.A., Croft, S.L., Nelson, D.J., 1985. Antileishmanial effect of allopurinol ribonucleoside and the related compounds, allopurinol, thiopurinol, thiopurinol ribonucleoside, and of formycin B, sinefungin and the lepidine WR6026. *Trans. R. Soc. Trop. Med. Hyg.* 79, 122–128.
- Neal, R.A., Iwobi, M.V., Robert-Gero, M., 1989. Antileishmanial effect of free and encapsulated sinefungin against *Leishmania donovani* infections in BALB/c mice. *C. R. Acad. Sci. Ser. III* 308, 485–488.
- Oketch-Rabah, H.A., Lemmich, E., Dossaji, S.F., Theander, T.G., Olsen, C.E., Cornett, C., Kharazmi, A., Christensen,

- S.B., 1997a. Two new antiprotozoal 5-methylcoumarins from *Vernonia brachycalyx*. *J. Nat. Prod.* 60, 458–461.
- Oketch-Rabah, H.A., Lemmich, E., Dossaji, S.F., Theander, T.G., Olsen, C.E., Cornett, C., Kharazmi, A., Christensen, S.B., 1997b. Antiprotozoal compounds from *Asparagus africanus*. *J. Nat. Prod.* 60, 1017–1023.
- Oketch-Rabah, H.A., Christensen, S.B., Frydenvang, K., Dossaji, S.F., Theander, T.G., Cornett, C., Watkins, W.M., Kharazmi, A., Lemmich, E., 1998. Antiprotozoal properties of 16,17-dihydrobrachycalyxolide from *Vernonia brachycalyx*. *Planta Med.* 64, 559–562.
- Paolantonacci, P., Lawrence, F., Nolan, L.L., Robert-Gero, M., 1987. Inhibition of leishmanial DNA synthesis by sinefungin. *Biochem. Pharmacol.* 36, 2813–2820.
- Parnham, M.J., 1996. Benefit–risk assessment of the squeezed sap of the purple coneflower (*Echinacea purpurea*) for long-term oral immunostimulation. *Phytomedicine* 3, 95–102.
- Patz, J.A., Graczyk, T.K., Geller, N., Vittor, A.Y., 2000. Effects of environmental change on emerging parasitic diseases. *Int. J. Parasitol.* 30, 1395–1405.
- Perez-Victoria, J.M., Tincusi, B.M., Jimenez, I.A., Bazzocchi, I.L., Gupta, M.P., Castans, S., Gamarro, F., Ravelo, A.G., 1999. New natural sesquiterpenes as modulators of daunomycin resistance in a multidrug-resistant *Leishmania tropica* line. *J. Med. Chem.* 42, 4388–4393.
- Pershin, G.N., Moskalenko, N.Y., 1963. Chemotherapeutic activity of monomycin in induced cutaneous leishmaniasis. *Farmakol. Toksikol. (Moscow)* 26, 97–101.
- Phelouzat, M.A., Basselin, M., Lawrence, F., Robert-Gero, M., 1995. Sinefungin shares adomet-uptake system to enter *Leishmania donovani* promastigotes. *Biochem. J.* 305, 133–137.
- Plock, A., Sokolowska-Köhler, W., Presber, W., 2001. Application of flow cytometry and microscopical methods to characterize the effect of herbal drugs on *Leishmania* spp. *Exp. Parasitol.* 97, 141–153.
- Queiroz, E.F., Roblot, F., Hocquemiller, R., Cavé, A., Barrios, A.A., Fournet, A., Ducrot, P.H., 1996. Pesseoine and spinosine, two catecholic berbines from *Annona spinescens*. *J. Nat. Prod.* 59, 438–440.
- Rasmussen, H.B., Christensen, S.B., Kvist, L.P., Karazmi, A., 2000a. A simple and efficient separation of the curcumins, the antiprotozoal constituents of *Curcuma longa*. *Planta Med.* 66, 393–398.
- Rasmussen, H.B., Christensen, S.B., Kvist, L.P., Kharazmi, A., Huansi, A.G., 2000b. Absolute configuration and antiprotozoal activity of minquartynoic acid. *J. Nat. Prod.* 63, 1295–1296.
- Rathore, A., Juneja, R.K., Tandon, J.S., 1989. An iridoid glucoside from *Nyctanthes arbor-tristis*. *Phytochemistry* 28, 1913–1917.
- Rezzano de Raffo, S., Moreno, G., Garcia, J., Scorza, J.V., 1985. Efficacy of paromomycin-I against *Leishmania garnhami* in heterozygous albino mice. *Rev. Cubana Med. Trop.* 37, 55–65.
- Richomme, P., Godet, M.C., Foussard, F., Toupet, L., Sevenet, T., Bruneton, J., 1991. A novel leishmanicidal labdane from *Polyalthia macropoda*. *Planta Med.* 57, 552–554.
- Ridoux, O., Di Giorgio, C., Delmas, F., Elias, R., Mshvildadze, V., Dekanosidze, G., Kemertelidze, E., Balansard, G., Timon-David, P., 2001. *In vitro* antileishmanial activity of three saponins isolated from ivy, alpha-hederin, beta-hederin and hederacolchiside A1, in association with pentamidine and amphotericin B. *Phyther. Res.* 15, 298–301.
- Robert-Gero, M., Bachrach, U., Baatnagar, S., Polonsky, J., 1985. Inhibition *in vitro* of the growth of *Leishmania donovani* promastigotes by quassinoids. *C. R. Acad. Sci. Ser. II* 300, 803–806.
- Rossi, B.R., Torres-Santos, E.C., Santos, A.P.P.T., Almeida, A.P., Costa, S.S., Da-Silva, S.A.G., 2000. Treatment of cutaneous leishmaniasis with *Kalanchoe pinnata*: experimental and clinical data. *Phytomedicine* 7, 115.
- Roy, R., Bhattacharya, G., Siddiqui, K.A.I., Bhadra, R., 1990. Berberine derivatives as antileishmanial drugs. *Antimicrob. Agents Chemother.* 34, 918–921.
- Saez, J., Granados, H., Torres, B., Velez, I.D., Munoz, D., 1998. Leishmanicidal activity of *Annona aff. spraguei* seeds. *Fitoterapia* 69, 478–479.
- Sahpaz, S., Bories, C.H., Loiseau, P.M., Cortes, D., Hocquemiller, R., Lauerns, A., Cavé, A., 1994. Cytotoxic and antiparasitic activity from *Annona senegalensis* seeds. *Planta Med.* 60, 538–540.
- Sahpaz, S., Gonazalez, M.C., Hocquemiller, R., Zafra-Polo, M.C., Cortes, D., 1996. Annosenegalin and annogalene: two cytotoxic mono-tetrahydrofuran acetoinins from *Annona senegalensis* and *Annona cherimolia*. *Phytochemistry* 42, 103–107.
- Santo, W.R., Bernardo, R.R., Pecanha, L.M.T., Palatnik, M., Parente, J.P., Sousa, C.B.P., 1997. Haemolytic activities of plant saponins and adjuvants. Effect of *Periandra mediterranea* saponin on the humoral response to the fml antigen of *Leishmania donovani*. *Vaccine* 15, 1024–1029.
- Savornin, B.M., Elias, R., Diaz-Lanza, A.M., Balansard, G., Gasquet, M., Delmas, F., 1991. Saponins of the ivy plant, *Hedera helix*, and their leishmanicidal activity. *Planta Med.* 57, 260–262.
- Sauvain, M., Dedet, J.P., Kunesch, N., Poisson, J., Gantier, J.C., Gayral, P., Kunesch, G., 1993. *In vitro* and *in vivo* leishmanicidal activities of natural and synthetic quinoids. *Phyther. Res.* 7, 161–171.
- Sauvain, M., Dedet, J.P., Kunesch, N., Poisson, J., 1994. Isolation of flavans from the amazonian shrub *Faramea guianensis*. *J. Nat. Prod.* 57, 403–406.
- Sauvain, M., Kunesch, N., Poisson, J., Gantier, J.C., Gayral, P., Dedet, J.P., 1996. Isolation of leishmanicidal triterpenes and lignans from the amazonian liana *Dolioscarpus dentatus* (Dilleniaceae). *Phyther. Res.* 10, 1–4.
- Schlein, Y., 1994. Mortality of *Leishmania major* in *Phlebotomus papatasi* caused by plant feeding of the sand flies. *Am. J. Trop. Med. Hyg.* 50, 20–27.
- Schmeda-Hirschmann, G., Razmilic, I., Sauvain, M., Moretti, C., Ruiz, E., Balanza, E., Fournet, A., 1996. Antiprotozoal activity of jatrogrossidione from *Jatropha grossidentata* and jatrophone from *Jatropha isabellii*. *Phyther. Res.* 10, 375–378.
- Schmeda-Hirschmann, G., Astudillo, L., Bastida, J., Codina, C., De Arias, A.R., Ferreira, M.E., Inchausti, A., Yaluff,

- G., 2001. Cryptofolione derivatives from *Cryptocarya alba* fruits. *J. Pharm. Pharmacol.* 53, 563–567.
- Seneca, H., Henderson, E., Harvey, M., 1948. Effect of hyaluronidase and of hyaluronic acid on cultures of *Trypanosomes*, *Leishmania*, and *Ameba*. *Science* 108, 714–715.
- Singha, U.K., Guru, P.Y., Sen, A.B., Tandon, J.S., 1992. Antileishmanial activity of traditional plants against *Leishmania donovani* in golden hamsters. *Int. J. Pharmacol.* 30, 289–295.
- Sittie, A.A., Lemmich, E., Olsen, C.E., Hvidt, L., Kharazmi, A., Nkrumah, F.K., Christensen, S.B., 1999. Structure–activity studies: *in vitro* antileishmanial and antimalarial activities of anthraquinones from *Morinda lucida*. *Planta Med.* 65, 316–319.
- Staerk, D., Lemmich, E., Christensen, J., Kharazmil, A., Olsen, C.E., Jaroszewski, J.W., 2000. Leishmanicidal, antiplasmodial and cytotoxic activity of indole alkaloids from *Corynanthe pachyceras*. *Planta Med.* 66, 531–536.
- Tahir, A.E., Ibrahim, A.M., Satti, G.M.H., Theander, T.G., Kharazmi, A., Khslid, A.S., 1998. The potential antileishmanial activity of some Sudanese medicinal plants. *Phytother. Res.* 12, 576–579.
- Tandon, J.S., Srivastava, V., Guru, P.Y., 1991. Iridoids: a new class of leishmanicidal agents from *Nyctanthes arbortristis*. *J. Nat. Prod.* 54, 1102–1104.
- Torres-Santos, E.C.S., Moreira, D.L., Kaplan, A.M.C., Meirelles, M.N., Rossi-Bergmann, B., 1999. Selective effect of 2',6'-dihydroxy-4'-methoxychalcone isolated from *Piper aduncum* on *Leishmania amazonensis*. *Antimicrob. Agents Chemother.* 43, 1234–1241.
- Vennerstrom, J.L., Lovelace, J.K., Waits, V.B., Hanson, W.L., Klayman, D.L., 1990. Berberine derivatives as leishmanial drugs. *Antimicrob. Agents Chemother.* 34, 918–921.
- Villaescusa, L., Diaz-Lanza, A.M., Martin, T., Gasquet, M., Delmas, F., Balansard, G., 1996. Preliminary screening of antiprotozoal activity of *Jasonia glutinosa* aerial parts. *Int. J. Pharmacogn.* 34, 303–304.
- Villaescusa, L., Diaz-Lanza, A.M., Gasquet, M., Dlemas, F., Ollivier, E., Bernabe, M., Faure, R., Elisa, R., Galansard, G., 2000. Antiprotozoal activity of sesquiterpene from *Jasonia glutinosa*. *Pharm. Biol.* 38 (3), 176–180.
- Waechter, A.I., Ferreira, M.E., Fournet, A., Arias, A.R., Nakayama, H., Torres, S., Hocquemiller, R., Cavé, A., 1997. Experimental treatment of cutaneous leishmaniasis with argentilactone isolated from *Annona haematantha*. *Planta Med.* 63, 433–435.
- Waechter, A.I., Yaluff, G., Inchausti, A., Arias, A.R., Hocquemiller, R., Cavé, A., Fournet, A., 1998. Leishmanicidal and trypanocidal activities of acetogenins isolated from *Annona glauca*. *Phytother. Res.* 12, 541–544.
- Waechter, A.I., Cavé, A., Hocquemiller, R., Bories, C., Munoz, V., Fournet, A., 1999. Antiprotozoal activity of aporphine alkaloids isolated from *Unonopsis buchtienii* (Annonaceae). *Phytother. Res.* 13, 175–177.
- Weinman, D., 1943. Effects of gramicidin and tyrocidine on pathogenic protozoa and a spirochete. *Proc. Soc. Exp. Biol. Med.* 54, 38–40.
- Weniger, B., Robledo, S., Arango, G.J., Deharo, E., Aragon, R., Munoz, V., Callapa, J., Lobstein, A., Anton, R., 2001. Antiprotozoal activities of Colombian plants. *J. Ethnopharmacol.* 78, 193–200.
- Werbel, L.M., Worth, D.F., 1980. Antiparasitics agents. Protozoal disease. *Ann. Rep. Med. Chem.*, 120–129.
- World Health Organization (WHO), 2002. Programme for the surveillance and control of leishmaniasis (<http://www.who.int/emc/diseases/leish/index.html>). Accessed 04/02/2002.
- Yang, D.M., Liew, F.Y., 1993. Effects of qinghaosu (artemisinin) and its derivatives on experimental cutaneous leishmaniasis. *Parasitology* 106, 7–11.
- Zhai, L., Blom, J., Chen, M., Christensen, S.B., Kharazmi, A., 1995. The antileishmanial agent licochalcone A interferes with the function of parasite mitochondria. *Antimicrob. Agents Chemother.* 39, 2742–2748.