

MINI REVIEW

Functional properties of Okra *Abelmoschus esculentus* L. (Moench): traditional claims and scientific evidences

Anupam Roy¹✉, Shanker Lal Shrivastava¹ and Santi M. Mandal²

Abstract

Okra, *Abelmoschus esculentus* L. (Moench) is an important vegetable crop cultivated in tropical, subtropical and warm temperate regions around the world. Besides the nutritional benefit, the different parts of the plant are used extensively in traditional medicine (antidiabetic, antipyretic, diuretic, antispasmodic, etc) around the world. This review critically assesses the nutritional values, phytochemistry, preclinical pharmacological properties and the possible future application of the okra. Effort is made to correlate the traditional claims in the context of experimental evidences.

Keywords: okra; traditional use; antidiabetic; antihyperlipidemic; dysentery; diarrhoea.

Introduction

Okra, *Abelmoschus esculentus* L. (Moench) commonly known as ladies finger and in several other vernacular names is cultivated as an important vegetable crop in tropical, subtropical and warm temperate regions around the world with total trade estimated to over \$5 billion (Oyenuga, 1969; Chauhan, 1972; Lamont, 1999; Oyelade, Ade-Omowaye, & Adeomi, 2003; Siemonsma & Kouame, 2000; Ndunguru & Rajabu, 2004; NRC, 2006; Kumar *et al.*, 2010; Benchasri, 2012; Lim, 2012). Okra is an annual or perennial tall (around 2 meters) dicotyledonous plant

related to species such as cotton, cocoa and *Hibiscus*. The plant grows preferably in well-drained humus rich fertile soil in full sun with pH ranging from 6 to 6.7, but it can tolerate a wide range of soil types and pH from 5.5 to 8.0 (N. Jain, R. Jain, V. Jain, & S. Jain, 2012). The leaves are long-petioled, orbicular or orbicular-ovate around 10–20 cm long, broad and rough, palmately lobed with 5–7 lobes. Flowers of this plant are axillary and solitary, 4–8 cm in diameter having five white to yellow petals, often with a red or purple spot at the base of each petal. Fruit is elongated, 10 to 25 cm long, 1.5 to 3 cm in diameter, tapering to a blunt point and containing rows of rounded, and kidney shaped seeds (Fig. 1). Depending on the cultivar, fruits of Okra mature after 60-180 days of sowing (alternatively can also be counted 5-10 days after flowering of plant). Fruits are detached from the stacks by applying slight twist (Tindall, 1986). Irritating hairs are sometimes present on leaves, stems and on the fruit surface.

Immature fresh and green seed pods are consumed as vegetable. It offers mucilaginous consistency after cooking. Often the extract obtained from the fruit is added to different recipes like soups, stews and sauces to increase the consistency. The immature pods are also used in making pickle. The entire plant is edible and is used to have several food (Babu & Srinivasan, 1995; Madison, 2008; Lim, 2012; Jain *et al.*, 2012; Maramag, 2013) and non food applications (Camciuc, Deplagne, Vilarem, & Gaset, 1998). Okra leaves are to some extent edible and are used as salad when fresh or cooked for edible purposes as the greens of beets or dandelions.

Okra seeds are source of oil and protein. It can be used as non-caffeinated substitute for coffee. Okra seeds may be roasted and ground to form a caffeine-free substitute for coffee (Martin, 1982; Calisir, Ozcan, Haciseferogullari, & Yildiz, 2005). Okra seed powder is used as a substitute for aluminium salts in water purification (Vaidya & Nanoti, 1989). Okra root mucilage has almost the same chemical composition as that of medicinal plant common marshmallow *Althaea officinalis* (Tomoda, Shimuzu, &

Received: 15 May 2014 / Accepted revised version: 5 June 2014 /
Published online: 16 July 2014

© Horizon e-Publishing Group

CITATION

Roy, A., Shrivastava, S. L., & Mandal, S. M. (2014). Functional properties of Okra *Abelmoschus esculentus* L. (Moench): traditional claims and scientific evidences. *Plant Science Today*, 1(3), 121-130. <http://dx.doi.org/10.14719/pst.2014.1.3.63>

AUTHORS' AFFILIATION

¹ Agricultural & Food Engineering Department, Indian Institute of Technology Kharagpur, Kharagpur-721302.

² Vidyasagar University and on Lien Indian Institute of Technology Kharagpur, Kharagpur-721302.

✉ CORRESPONDENCE: Anupam Roy, Tel: +91-9734461153; E-mail: anupamroypaper@gmail.com

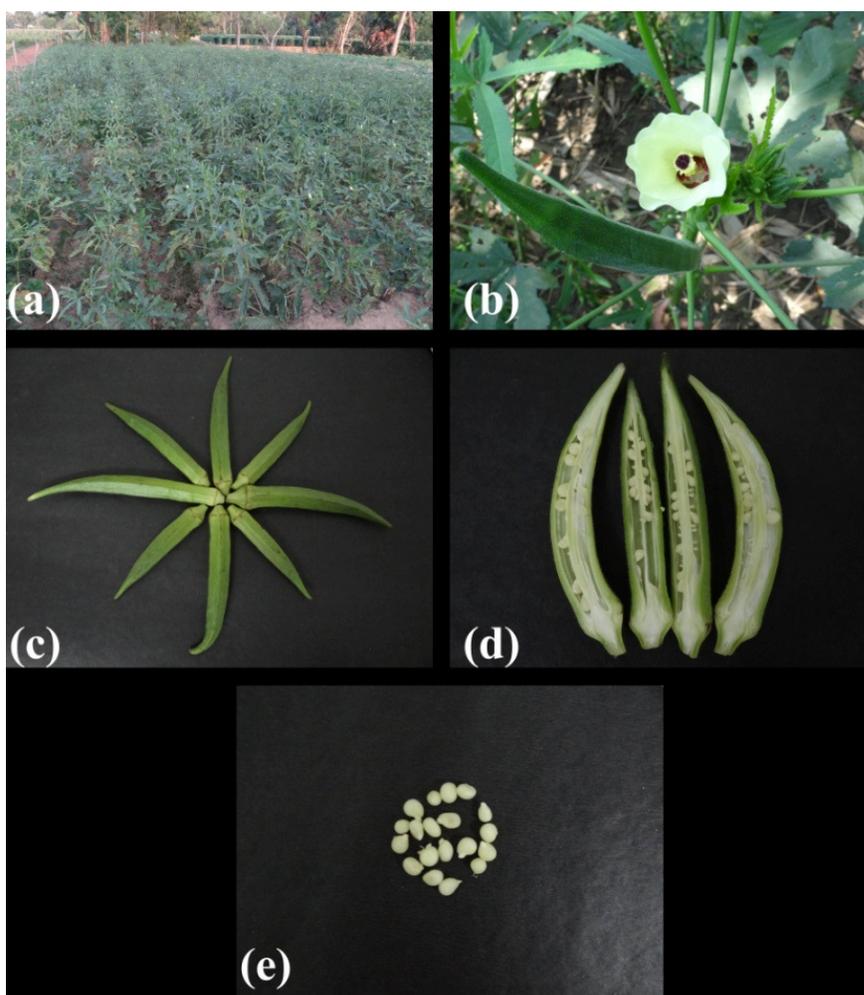


Fig 1. Different parts of okra (a) Okra in field (b) Okra fruit and flower (c) Okra fruit (d) Cross sectional view (e) Okra seed.

Gonda, 1985).

Other non-food applications include use of the root and stem of okra for cleaning the cane juice from which Jaggery (gur or brown sugar) is prepared (Chauhan, 1972). Mature fruits and stems containing crude fiber are used in the paper industry (Martin, 1982). It can also be used for sacks and ropes (Watt, 1908), biogas and fuel (Dahiya & Vasudevan, 1987).

Okra is widely used in ethno medicine in diverse cultures (Table 1). In Ayurveda, okra is used as an edible infusion and in different preparation for diuretic effect (Maramag, 2013). An infusion of the fruit mucilage is also used to treat dysentery and diarrhoea in acute inflammation and irritation of the stomach, bowels, and kidneys catarrhal infections, ardour urinae, dysuria and gonorrhoea. Seeds are used as antispasmodic, cordial and stimulant (Lim, 2012, pp. 160-169). Leaves and root extracts are served as demulcent and emollient poultice (Babu & Srinivasan, 1995).

Although some reviews are available describing

medicinal properties okra (Kumar *et al.*, 2010; Indah Mohd, 2011; Fong, Toh, Rajen, & Rao, 2011; Jain *et al.*, 2012; Nwachukwu, Nulit & Rusea, 2014), but no specific review are present describing nutritional values, phytochemistry, preclinical pharmacological properties and the possible future application of the okra. This review is focused to draw a correlation between traditional ethno-medicinal claims of okra with the established scientific evidences. Besides discussion are drawn on the future prospect on the different ways of applying this nutraceutical in food and medicinal matrices.

Nutritive and phytochemical profiling of different parts of okra (*A. esculentus*)

Okra pods are mucilaginous, low in calories but nutritionally rich and a good source of edible fiber. Studies have shown that the okra pod contains important bioactive compounds such as carotene, folic acid, thiamine, riboflavin, niacin, vitamin C, oxalic acid and amino acids.

Pods are low in saturated fat, very low in cholesterol and offers sufficient amount of minerals (Table 2) (U.S

Table 1. Okra in Ethnomedicine

| Parts | Form | Name of the Medicinal system where it is used | Used for | References |
|--------|---|---|--|---|
| Fruit | Infusion of the fruit mucilage | Indian ethnomedicine | For treating dysentery and diarrhoea in acute inflammation and irritation of the stomach, bowels, and kidneys catarrhal infections, ardour urinae, dysuria, diuretic, plasma replacement and gonorrhoea. | Odedra, & Nathabhai, 2009; Lim, 2012; Maramag, 2013; Smit, Neeraj, & Preeti, 2013; Sayana <i>et al.</i> , 2014 |
| | Infusion of the fruit mucilage | Indian ethnomedicine | Antipyretic and plasma replacement. | |
| | A decoction of the immature fruit | Indian ethnomedicine | Demulcent and emollient poultice. | |
| Leaves | Extract of leaves and roots | Indian ethnomedicine | Demulcent, though less so than that of okra fruit. | Babu & Srinivasan, 1995; Odedra, & Nathabhai, 2009 |
| | Extract of leaves | Indian ethnomedicine | Extract of leaves mixed with egg albumin and applied on hair which makes black and silky hair. | |
| | Leaves | Latin America | Remedies for tumour | |
| Root | Extract of roots | Indian ethnomedicine | Demulcent and emollient poultice. | Barrett, 1994; Yesilada <i>et al.</i> , 1951; Babu, & Srinivasan, 1995; Odedra, & Nathabhai, 2009; Lim, 2012 |
| | The juice of the roots | Nepal | To treat cuts, wounds and boils. | |
| | An infusion of the roots | Indian ethnomedicine, Malaya | Treatment of syphilis. | |
| | Infusion of the roots | Traditional medicine of Nicaragua's Atlantic Coast and Turkey | Used as stomachic, to treat diabetes, ulcer, used as laxative and treatment of jaundice. | |
| Seed | Seeds | Indian ethnomedicine | Antispasmodic, cordial and stimulant. | Crossley & Hilditch, 1952; Martin, 1982; Vaidya & Nanoti, 1989; Calisir <i>et al.</i> , 2005; Jarret <i>et al.</i> , 2011; Lim, 2012 pp. 160-167; Aslan, Sezik, Yesilada, 2003; Smit <i>et al.</i> , 2013 |
| | Infusion of the roasted seeds | Indian ethnomedicine | Has sudorific properties | |
| | Okra seed | Indian ethnomedicine | Treatment of spermatorrhoea | |
| | Okra seed | Turkish folk medicine | In managing increased blood glucose concentration | |
| | Seeds | Latin America | Remedies for tumour | |
| | Infusion of roasted okra seeds | Turkey | Diabetes mellitus therapy. | |
| Flower | The decoction of the leaves and flowers | Indian ethnomedicine | Used for the treatment of bronchitis and pneumonia. | Lim, 2012; Marwat <i>et al.</i> , 2011. |

Department of Agriculture, Agricultural Research Service 2010, Lim, 2012).

Random coil polysaccharides consisting of galactose, rhamnose, and galacturonic acid are in the structural form

of okra gum (Fig. 2). α (1-2)-rhamnose and α (1-4)-galacturonic acid residues with disaccharide side chains and a degree of acetylation (DA = 58) are the repeating unit of gum (Alamri, Mohamed, Hussain, & Xu,

2012; Zaharuddin, Noordin, & Kadivar, 2014). Besides this plant is also the treasure house of polyphenolic compounds. Presence of hyperoside, quercetin, coumarin scopoletin, uridine, and phenylalanine is reported by several authors (Bandyukova & Ligai, 1987; Lu, Huanfen, & Linlin, 2011). Shui & Peng (2004) have reported that quercetin derivatives and (-)-epigallocatechin as major antioxidant compounds in okra. 70% of the total antioxidant activity comes due to the quercetin derivatives (quercetin 3-O-xylosyl (1"→2") glucoside, quercetin 3-O-glucosyl (1"→6") glucoside, quercetin 3-O-glucoside and quercetin 3-O- (6"-O-malonyl)- glucoside). Liao, Dong, Shi, Liu, & Yuan, (2012) reported a new flavonol glycoside named 5, 7, 3', 4'-tetrahydroxy-4"-O-methyl flavonol-3-O-β-D- glucopyranoside with another pre reported compound 5, 7, 3', 4'-tetrahydroxy flavonol-3-O-[β-D-glucopyranosyl- (1→6)]-β-D-glucopyranoside.

Table 2. The proximate value per 100g edible portion of okra

| Item | Quantity |
|---------------------|------------------|
| Water | 90.17 g |
| Energy | 31 kcal (129 kJ) |
| Protein | 2.00 g |
| Total lipid | 0.10 g |
| Ash | 0.70 g |
| Carbohydrate | 7.03g |
| Total dietary fibre | 3.2 g |
| Total sugars | 1.2 g |
| Sucrose | 0.40 g |
| Glucose | 0.13 g |
| Fructose | 0.21 g |
| Starch | 0.34 g |
| Minerals | |
| Ca | 81 mg |
| Fe | 0.8 mg |
| Mg | 57 mg |
| P | 63 mg |
| K | 303 mg |
| Na | 8 g |
| Zn | 0.60 mg |
| Cu | 0.094 mg |
| Mn | 0.990 mg, |
| Se | 0.7 m g |
| Amino acids | |
| Tryptophan | 0.017 g |
| Threonine | 0.065 g |
| Isoleucine | 0.069 g |
| leucine | 0.105 g |
| lysine | 0.081 g |
| methionine | 0.021 g |
| Cystine | 0.019 g |

| | |
|---------------------|---------|
| Phenylalanine | 0.065 g |
| Tyrosine | 0.087 g |
| Valine | 0.091 g |
| Histidine | 0.031 g |
| alanine | 0.073 g |
| Aspartic acid | 0.145 g |
| Glutamic acid | 0.271 g |
| Glycine | 0.044 g |
| Proline | 0.045 g |
| Serine | 0.044 g |
| Lutein + zeaxanthin | 516 m g |
| Arginine | 0.084 g |

| Lipid | |
|---|------------|
| Total Saturated fatty acids | 0.026 g |
| Palmitic acid (16:0) | 0.022 g |
| Stearic acid (18:0) | 0.003 g |
| Total mono-unsaturated fatty acids | 0.017 g |
| Oleic acid (18:1) | 0.016 g |
| Total polyunsaturated fatty acids | 0.027 g, |
| Undifferentiated (linoleic acid; 18:2) | 0.026 g |
| Undifferentiated (linolenic acid; 18:3) | 0.001 g |
| Phytosterols | 24 mg |
| Vitamin | |
| Vitamin C | 21.1 mg |
| Thiamine | 0.02 mg |
| Riboflavin | 0.060 mg |
| Niacin | 1.0 mg |
| Pantothenic acid | 0.245 mg |
| Vitamin B- | 6 0.215 mg |
| Total folate | 88 m g |
| Total choline | 12.3 mg |
| β-carotene | 225 m g |
| Vitamin A | 375 IU |
| Vitamin A RAE | 19 m g |
| Vitamin E (a-tocopherol) | 0.36 mg |
| Vitamin K (phylloquinone) | 53 m g |

Mature seeds are used for oil production and, when ground, as a substitute for coffee. Whole seeds and kernels of okra are rich in protein as well as fat. Major portion of protein and fat of the seed is accumulated in the kernel while crude fibre is concentrated in the seed coat or hull. It is rich in essential amino acids, has trypsin activity and chemical score of 67 (Rao, 1985). Okra seed oil is rich in palmitic, oleic, and linoleic acids (Crossley & Hilditch, 1951; Chisholm, & Hopkins, 1957; Steyn *et al.*, 2014). The value of palmitic acid in *A. esculentus* was in the range of 10.3%-36.35%, where as values of linoleic acid was from 23.6 to 50.65% (Jarret, Wang, & Levy, 2011).

Root mucilage of this plant poses similar chemical composition as that of medicinal plant *Althaea officinalis*. It was composed of partially acetylated acidic polysaccharide

(molecular weight about 1700000 Da. The polysaccharide was composed of L-rhamnose : D-galactose : D-galacturonic acid : D-glucuronic acid : O-acetyl groups in the molar ratio of 1.1 : 1.9 : 1.0 : 1.0 : 2.0 (Tomoda, Shimizu, & Gonda, 1985). The phytochemical study of the root extract confirmed presence of carbohydrate, fixed oils, mucilage and flavanoid glycosides (Tomoda *et al.*, 1987) and offers antioxidant potential (Sunilson, Jayaraj, Mohan, Kumari, Varatharajan, 2008).

According to the analytical study of the leaves of *A. esculentus* (Nwachukwu *et al.*, 2014, pp. 16-19; Idris, Yisa, & Itodo, 2009), the proximate and mineral composition were 82.53±1.60% moisture, 18.48±0.03% ash, 7.63±0.06% crude protein, 12.98±0.03% crude lipid and 27.54±0.27% available carbohydrate. Dominant mineral elements found in leaves were K (2107.50±0.03 mg/100g) and Mg (75.85±0.02 mg/100mg) where as appreciable concentrations of Na (37.50±0.83 mg/100g), Ca (57.03±0.12 mg/100g), P (7.33±0.04 mg/100g), Cu (3.22±0.02 mg/100g), Fe (20.78±0.15 mg/100g), Mn (17.25±0.22 mg/100g) and Zn (8.64±0.04 mg/100g) were also found. Hedin, Lamar, Thompson, & Minyard (1968) identified 11 flavonol glycosides *i.e.* quercetin 4'-glucoside, quercetin 7-glucoside, quercetin 5-glucoside, quercetin 3-diglucoside, quercetin 4'-diglucoside, quercetin 3-triglucoside, quercetin 5-rhamnoglucoside, gossypetin 8-glucoside, gossypetin 8-rhamnoglucoside, gossypetin 3-glucosido-8-rhamnoglucoside, and the anthocyanins were cyanidin 4'-glucoside and cyaniding 3-glucosido-4' glucoside and two anthocyanins from the flower petal of this plant.

Validated pharmacological properties of the okra

Antioxidant activity and prevention of cellular damage related diseases

Reactive oxygen species (ROS) *i.e.* superoxide anion (O₂⁻), hydrogen peroxide (H₂O₂), and the hydroxyl radical (OH⁻) and reactive nitrogen species (RNS) *i.e.* nitric oxide (NO), peroxyxynitrite (ONOO⁻) when produced in excess, cause cell dysfunction and ultimately death. This happens due to alteration of metabolic pathway activity (Newsholme, Keane, Welters, & Morgan, 2007; Newsholme *et al.*, 2009) and/or the structure of cellular membranes, DNA, or proteins (Chandra, Samali, & Orrenius, 2000; Limon-Pacheco, & Gonsebatt, 2009; Newsholme *et al.*, 2012). Many medicinal plants, fruits and their products, fermented food, etc are proved to have sufficient antioxidant to scavenge these free radicals and to prevent the ensuing damage (Sánchez-Moreno, Larrauri, & Saura-Calixto, 1999; Alia *et al.*, 2008; Krishnaiah, Sarbatly, & Nithyanandam, 2011; Roy *et al.*, 2012a; Roy, Khanra, Mishra, & Bhattacharyya, 2012b; Roy, Khanra, Mishra, Bhattacharya, & Bhattacharyya, 2012c; Zhou, Zhang, Sun, Yan, & Wang, 2014).

With regard to Okra, several studies have been conducted on the antioxidant activity with different parts of the plant. Atawodi *et al.* (2009) has reported *in vitro* antioxidant assay of methanol extract of okra fruits. They have done antioxidant/radical scavenging activities by xanthine oxidase and 2-deoxyguanosine methods and reported 50% inhibitory concentration values of 25 and 43 ml. According to Khomsug, Thongjaroenbuangam, Pakdeenarong, Suttajit, & Chantiratikul (2010), total phenolic content of pulped and seeds of okra extracts as 10.75±0.02mg GAE/100g extract and 142.48±0.02mg GAE/100g extract which corresponds with scavenging activities. Besides they have also found procyanidin B2 as predominant phenolic compound followed by procyanidin B1 and rutin in seeds. In pulped seed catechin, procyanidin B2, epicatechin and rutin are reported to be present. It is quite important to see that roasting (1600°C for 10–60 minutes) increased the nutrient composition and antioxidant activity of the seeds (Adelakun, Ade-Omowaye, Adeyemi, & Van De Venter, 2010) whereas pre-treatment (soaking and blanching) increased the nutrient composition, but decreases antioxidant activity (Adelakun *et al.*, 2009). Ansari, Houlihan, Hussain, & Pieroni (2005) reported Okra extract as *in vitro* non-enzymatic inhibitor of lipid peroxidation in liposomes. *A. esculentus* peel and seed powder contains significant *in vivo* antioxidant property in streptozotocin-induced diabetic rats.

Administration of different doses of peel and seed powder significantly increased liver, kidney and pancreas superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPx), reduced glutathione (GSH) levels and decreased thiobarbituric acid reactive substances (TBARS) (P < 0.001) levels in diabetic rats compared to diabetic control rats. Liao, Liu, & Yuan, (2012) has done a comparative analysis of total phenolics and total flavonoids and antioxidant ability of different organs (flower, fruit, leaf, and seed) and different enrichment fractions of water extracts of the *A. esculentus* plant. They confirmed fruitful presence of total phenolics and total flavonoids related to antioxidant ability in all the extracts of the plant organs although percentage varied. In flower of okra highest amount of total phenolics and total flavonoids were found (Liao *et al.*, 2012).

This data suggests Okra as a good contributor to the antioxidant status and promising chemopreventive agent as described in several traditional medicines for human race.

Okra as antidiabetic and antihyperlipidemic and related disease prevention

In traditional medicine Okra seeds are reported to have ability in managing increased blood glucose concentration. Modern research has correlated this traditional claim with

scientific evidences.

Tomoda *et al.* (1989) reported that okra polysaccharide possesses anticomplementary and hypoglycemic activity in normal mice. *A. esculentus* was found to have hypolipidemic activity in *in vivo* tested rat model (Trinh, Nguyen, Tran, & Nguyen 2008) and in mice (Ngoc, Ngo, Van, & Phung, 2008). Okra polysaccharide lowers the cholesterol level in blood and may prevent cancer by its ability to bind bile acids (Kahlon, Chapman & Smith, 2007).

Cholesterol levels decreased 56.45%, 55.65%, 41.13%, 40.50% and 53.63% respectively in mice groups orally administered with dichloromethane okra plant extract, methanol okra plant extract, dichloromethane okra fruit extract, methanol okra fruit extract and simvastatin as compared to the tyloxapol injected group (Ngoc *et al.*, 2008). The effects of crude extracts of *A. esculentus* on albumin and total bilirubin levels of diabetic albino rats were reported to have a significant ($P < 0.05$) increase (82%) in total bilirubin levels in diabetic control group over the normal control (Uraku, Ajah, Okak, Ibiem, & Onu, 2010). Ramachandran, Sandeep, Srinivas, & Dhanaraju, 2010 reported anti-diabetic activity of okra on alloxan-induced diabetic rats. Sabitha, Ramachandran, Naveen, & Panneerselvam Sabita *et al.*, (2012, 2013) has reported antidiabetic and antihyperlipidemic potential of okra peel and seed powder in streptozotocin (STZ)-induced diabetic rats. Administration of peel and seed powder at 100 and 200 mg/kg dose in diabetic rats showed significant ($P < 0.001$) reduction in blood glucose level and increase in body weight than diabetic control rats. Water-soluble fraction of the fruits of Okra was studied to check the absorption of oral glucose as well as metformin from the gastrointestinal tract in the Long Evans rats. It showed significant reduction in absorption of glucose as studied in the 24 hours fasting rats (Khatun, Rahman, Biswas, & Islam, 2011). Thanakosai & Phuwapraisirisan, (2013) has reported, the presence of two major flavonol glucosides named isoquercetin (2) and quercetin-3-O-beta-glucopyranosyl- (1"→6")-glucoside (3) in okra seeds which are α -glucosidase inhibitors. These two compounds selectively inhibited rat intestinal maltase and sucrase, in which isoquercetin (2) was 6-10 times more potent than its related diglucoside 3. Subrahmanyam *et al.*, (2011) has reported antidiabetic activity of okra fruit extract.

The effects of *A. esculentus* fruits on alkaline phosphatase (ALP), aspartate aminotransferase (AST) and alanine aminotransferase (ALT) activities on diabetic albino rats were also investigated. Serum glucose levels and activities of enzymes *viz.* ALP, AST and ALT decreased significantly after administration of the extracts (Uraku, Onuoha, Offor, Ogbanshi, & Ndidi, 2011). Hypoglycemic effect of ethanolic and aqueous extract of *A. esculentus* fruit was studied. Results revealed that aqueous extract of

powdered drug had maximum effect (Saha, Jain, & Jain, 2011). Recent study reported that the extract of okra lowers blood glucose and serum lipids in high-fat diet-induced obese C57BL/6 mice. Ethanol extract of okra (EO) and its major flavonoids isoquercitrin and quercetin 3-O-gentiobioside reduced blood glucose and serum insulin levels and improved glucose tolerance in obese mice (Fan *et al.*, 2014).

For treating dysentery and diarrhoea in acute inflammation and irritation of the stomach, bowels

In Asia and African traditional medicine, okra fruits are served as mucilaginous food as a dietary meal in the treatment of gastric irritations and inflammative diseases. Scientific explanation of such use came in recent years. Lengsfeld, Titgemeyer, Faller, & Hensel (2004) pre-treated *Helicobacter pylori* with a fresh juice of okra that completely inhibited adhesion in an *in situ* adhesion model on sections of human gastric mucosa. The anti-adhesive qualities of okra were assumed to be due to a combination of glycoproteins and highly acidic sugar compounds making up a complex three-dimensional structure that is fully developed only in the fresh juice of the fruit. That is due to the blocking capacity of specific *Helicobacter* surface receptors that coordinate the interaction between host and bacterium. According to Messing *et al.*, 2014, it supported the previous claims and showed that the effectiveness in treating gastric irritations and inflammative diseases is due to polysaccharides that inhibit the adhesion of *H. pylori* to stomach tissue.

Recent trends and future prospect

Okra extract is used as a key ingredient in several commercially important products of food and medicine. The rheological behaviour (Kontogiorgosa, Margeloua, Georgiadisb, & Ritzoulisb 2013), properties of forming oil water emulsion (Georgiadisa *et al.*, 2011) and ability to stabilize acidic emulsion (Alba, Ritzoulis, Georgiadis, & Kontogiorgos 2013) of okra can potentially be used as future value addition applications like composite materials (Dimopoulou & Ritzoulis, 2014) and food foam productions (Laporte *et al.*, 2014) .

In last decade, extensive efforts have been given in developing of several nanoscale-carriers in to improve the drug delivery systems (Roy *et al.*, 2012a, 2012b; Mandal *et al.*, 2014). Okra may play a leading role in improved drug delivery system. Several reports came using okra polysaccharide as drug release agent. Okra gum as a mini-matrix for furosemide and diclofenac sodium tablets showed prolonged release of furosemide and diclofenac sodium from the compressed tablets (Ofoefule & Chukwu, 2001). Besides it is now used as a medium for several other drug deliveries. Bakre and Jaiyeoba (2009) used it as metronidazole tablet formulation. Sharma, Kulkarni, & Sharma, (2013a) used it in the development mucoadhesive

gel for nasal delivery of rizatriptan benzoate. Recently this same research group (Sharma, Kulkarni, Sharma, Bhatnagar, & Kumar, 2013b) have prepared and evaluated of mucoadhesive microspheres, using okra polysaccharide as a novel carrier for safe and effective delivery of rizatriptan benzoate into nasal cavity. It is also used to study the sustaining release of drug (Zaharuddin *et al.*, 2014). Besides colon specific drug delivery studies also been carried out (Rajkumari, Sarma, Ilango, Devi, & Rajak, 2012). If drug release is the present hunk of okra research, the future might come as a medium of probiotic, nutraceutical delivery. Several new formulation might come like edible coating, preservative carrier etc. So more application oriented research might be carried out to get the full utilization of this novel natural gift.

Conclusion

From previous discussion it can be concluded that traditional ethnomedicinal claims of Okra (*A. esculentus*) has a strong scientific evidences. Traditionally claimed pharmacological properties of okra attributed to the presence of various phyto-compounds reported. The strong scientific evidence of *in vitro* and *in vivo* biological activity confirms the doubt of its traditional use. Detailed investigations for its myriad beneficial effects may enlighten the future of medicinal exploitation. However further research should be focused to find out the mechanism of action of the pharmacological activities at the molecular level. This can solve several unanswered questions of origin, development and cure of diseases. Besides, being nontoxic in nature, this fruit can be easily tried for human trials rather than animal models. Okra based anti-diabetic food, antioxidant rich food formulation can be thus easily be tried avoiding complicated medical trials. It would get go for better value addition and commercialization in near future not being confined only in kitchen.

Acknowledgments

We are thankful to CSIR, India for providing CSIR Individual Fellowship to Anupam Roy (CSIR sanction No- 9/1103 (0001)2k13-EMR-I).

References

Adelakun, O. E., Ade-Omowaye, B. I. O., Adeyemi, I. A., & Van De Venter, M. (2010). Functional properties and mineral contents of a Nigerian okra seed (*Abelmoschus esculentus* Moench) fl our as influenced by pretreatment. *Journal of Food Technology*, 8(2), 39–45. <http://dx.doi.org/10.3923/jftech.2010.39.45>

Adelakun, O. E., Oyelade, O. J., Ade-Omowaye, B. I., Adeyemi, I. A., Van de Venter, M., & Koekemoer, T. C. (2009). Influence of pre-treatment on yield chemical and antioxidant properties of a Nigerian okra seed (*Abelmoschus esculentus* Moench) fl our. *Food and Chemical*

Toxicology, 47 (3), 657–661. PMID:19146911. <http://dx.doi.org/10.1016/j.fct.2008.12.023>

- Alamri, M. S., Mohamed, A., Hussain, S., & Xu, J. (2012). Effect of Okra extract on properties of wheat, corn and rice starches, *Journal of Food, Agriculture and Environment*, vol. 10(1), 217–222.
- Alba, K., Ritzoulis, C., Georgiadis, N., & Kontogiorgos, V. (2013). Okra extracts as emulsifiers for acidic emulsions. *Food Research International*, 54(2), 1730–1737. <http://dx.doi.org/10.1016/j.foodres.2013.09.051>
- Alia, S. S., Kasojua, N., Luthraa, A., Singha, A., Sharanabasavaa, H., Sahu, A., & Bora, U. (2008). Indian medicinal herbs as sources of antioxidants. *Food Research International*, 41, 1–15. <http://dx.doi.org/10.1016/j.foodres.2007.10.001>
- Ansari, N. M., Houlihan, L., Hussain, B., & Pieroni, A. (2005). Antioxidant activity of five vegetables traditionally consumed by south-Asian migrants in Bradford, Yorkshire, UK. *Phytotherapy Research*, 19(10), 907–911. <http://dx.doi.org/10.1002/ptr.1756> PMID:16261524
- Atawodi, S. E., Atawodi, J. C., Idakwo, G. A., Pfundstein, B., Haubner, R., Wurtele, G., Spiegelhalder, ... Owen, R. W. (2009). Polyphenol composition and antioxidant potential of *Hibiscus esculentus* L. fruit cultivated in Nigeria. *Journal of Medicinal Food*, 12(6), 1316–1320. <http://dx.doi.org/10.1089/jmf.2008.0211> PMID:20041787
- Babu, P. S., & Srinivasan, K. (1995). Influence of dietary curcumin and cholesterol on the progression of experimentally induced diabetes in albino rat. *Molecular and Cellular Biochemistry*, 152, 13–21. PMID:8609907
- Bakre, L. G., & Jaiyeoba, K. T. (2009). Effects of drying methods on the physicochemical and compressional characteristics of Okra powder and the release properties of its metronidazole tablet formulation. *Archives of Pharmacal Research*, 32 (2), 259–67.
- Bandyukova V. A., & Ligai, L. V. (1987). A chemical investigation of the fruit of *Abelmoschus esculentus*, *Chemistry of natural compounds*, 23, 376–7. <http://dx.doi.org/10.100/BF00600851>
- Barrett, B. (1994). Medicinal plants of Nicaragua's Atlantic Coast. *Economic Botany*, 48(1), 8–20. <http://dx.doi.org/10.1007/BF02901375>
- Benchasri, S. (2012). Okra (*Abelmoschus esculentus* (L.) Moench) as a Valuable Vegetable of the World Ratar. *Ratarstvo i povrtarstvo*, 49, 105–112.
- Calisir, S., Ozcan, M., Haciseferogullari, H., & Yildiz, M. U. (2005). A study on some physico-chemical properties of Turkey okra (*Hibiscus esculenta*) seeds. *Journal of Food Engineering*, 68, 73–78. <http://dx.doi.org/10.1016/j.jfoodeng.2004.05.023>
- Camciuc, M., Deplagne, M., Vilarem, G., & Gaset, A. (1998). Okra - *Abelmoschus esculentus* L. (Moench.) a crop with economic potential for set aside acreage in France. *Industrial Crops and Products*, 7, 257–264. [http://dx.doi.org/10.1016/S0926-6690\(97\)00056-3](http://dx.doi.org/10.1016/S0926-6690(97)00056-3)
- Chandra, J., Samali, A., & Orrenius, S. (2000). Triggering and modulation of apoptosis by oxidative stress. *Free Radical Biology & Medicine*, 29, 323–333. [http://dx.doi.org/10.1016/S0891-5849\(00\)00302-6](http://dx.doi.org/10.1016/S0891-5849(00)00302-6)
- Chauhan, D. V. S. (1972). *Vegetable Production in India*, 3rd ed., Agra, India: Ram Prasad and Sons.

- Chisholm, M. J. & Hopkins, C. Y. (1957). An oxygenated fatty acid from the seeds of *Hibiscus esculentus*. *Canadian Journal of Chemistry*, 35, 358–364.
- Crossley, A., & Hilditch, T. P. (1951). The fatty acids and glycerides of okra seed oil. *Journal of the Science of Food and Agriculture*, 2, 251–255.
- Dahiya, A.K., & Vasudevan, P. (1987). Farm biomass utilization alternatives. *Biological Wastes*, 21, 85–91. [http://dx.doi.org/10.1016/0269-7483\(87\)90132-7](http://dx.doi.org/10.1016/0269-7483(87)90132-7)
- Dimopoulou, M., & Ritzoulis, M. (2014). Composite materials based on okra hydrocolloids and hydroxyapatite. *Food Hydrocolloids*. <http://dx.doi.org/10.1016/j.foodhyd.2014.04.015>
- Fan, S., Zhang, Y., Sun, Q., Yu, L., Li, M., ... Huang, C. (2014). Extract of okra lowers blood glucose and serum lipids in high-fat diet-induced obese C57BL/6 mice. *The Journal of Nutritional Biochemistry*. <http://dx.doi.org/10.1016/j.jnutbio.2014.02.010>
- Fong, Y., Toh, J., Rajen, M., & Rao, A. N. (2011). Recent researches on okra: *Abelmoschus esculentus*. *Journal of Tropical Medicinal Plants*, 12(1), 95-98.
- Georgiadisa, N., Ritzoulisa, C., Siouraa, G., Kornezoua, P., Vasiliadoub, C., & Tsiopstiasa, C. (2011). Contribution of okra extracts to the stability and rheology of oil-in-water emulsions. *Food Hydrocolloids*, 25(5), 991–999.
- Hedin, P. A., Lamar, P. L., Thompson, A. C., & Minyard, J. P. (1968). Isolation and structural determination of 13 flavonoid glycosides in *Hibiscus esculentus* (okra). *American Journal of Botany*, 55 (4), 431–437 <http://dx.doi.org/10.2307/2440572> PMID:5641298
- Idris, S., Yisa, J., & Itodo, A. (2009) Proximate and mineral composition of the leaves of *Abelmoschus esculentus*. *International Journal of Tropical Agriculture and Food Systems*, 3(2). <http://dx.doi.org/10.4314/ijotafsv3i2.50037>
- Indah Mohd, I. (2011). Amin Nutritional Properties of *Abelmoschus Esculentus* as Remedy to Manage Diabetes Mellitus: A Literature Review. *International Proceedings of Chemical, Biological and Environmental Engineering*, 11. Singapore: IACSIT Press.
- Jain, N., Jain, R., Jain, V., & Jain, S. (2012). A review on: *Abelmoschus esculentus*. *Pharmacia*, 1 (3), 84-89.
- Jarret, R. L, Wang, M. L, & Levy, I. J. (2011). Seed oil and fatty acid content in okra (*Abelmoschus esculentus*) and related species. *Journal of Agricultural Food Chemistry*, 59(8), 4019-24. <http://dx.doi.org/10.1021/jf104590u>
- Kahlon, T. S., Chapman, M. H., & Smith, G. E. (2007). *In vitro* binding of bile acids by okra, beets, asparagus, eggplant, turnips, green beans, carrot and cauliflower. *Food Chemistry*. 103, 676–80. <http://dx.doi.org/10.1016/j.foodchem.2006.07.056>
- Khatun, H., Rahman, A., Biswas, M., & Islam, A. U. (2011). Water-soluble Fraction of *Abelmoschus esculentus* L Interacts with Glucose and Metformin Hydrochloride and Alters Their Absorption Kinetics after Coadministration in Rats. *ISRN Pharmaceutical*, 260537. <http://dx.doi.org/10.5402/2011/260537>
- Khomsug, P., Thongjaroenbuangam, W., Pakdeenarong, N., Suttajit, M., & Chantiratikul P. (2010). Antioxidative Activities and Phenolic Content of Extracts from Okra (*Abelmoschus esculentus* L.) *Research Journal of Biological Sciences*, 5(4), 310-313 <http://dx.doi.org/10.3923/rjbsci.2010.310.313>
- Kontogiorgosa, V., Margeloua, I., Georgiadisb, N., & Ritzoulisb, C. (2012). Rheological characterization of okra pectins. *Food Hydrocolloids*, 29(2), 356–362
- Krishnaiah, D., Sarbatly., R, & Nithyanandam, R. (2011). A review of the antioxidant potential of medicinal plant species. *Food and Bioproducts Processing*, 89(3), 217–233. <http://dx.doi.org/10.1016/j.fbp.2010.04.008>
- Kumar, S., Dagnoko, S., Haougui, A., Ratnadass, A., Pasternak, D., & Kouame, C. (2010). Okra (*Abelmoschus pp.*) in West and Central Africa: potential and progress on its improvement. *African Journal of Agricultural Research*, 5, 3590-3598.
- Lamont, W. (1999). Okra a versatile vegetable crop. *Horticulture Technology* 9, 179-184.
- Laporte, M., Della, Valle, D., Loisel, C., Marze, S., Riaubanc, A., & Montillet, A. (2014). Rheological Properties of Food Foams Produced by SMX Static Mixers. *Food Hydrocolloids*. Article In Press, Accepted Manuscript.
- Lengsfeld, C., Titgemeyer, F., Faller, G., & Hensel, A. (2004). Glycosylated compounds from okra inhibit adhesion of *Helicobacter pylori* to human gastric mucosa. *Journal of Agricultural Food Chemistry*, 52(6), 1495-503. <http://dx.doi.org/10.1021/jf030666n> PMID:15030201
- Liao, H, Liu, H., Yuan, K. (2012). A new flavonol glycoside from the *Abelmoschus esculentus* Linn. *Pharmagnosy Magazine*, 8, 12-5. <http://dx.doi.org/10.4103/0973-1296.93303> PMID:22438657 PMID:PMC3307196
- Liao, H., Dong, W., Shi, X., Liu, H., & Yuan, K. (2012). Analysis and comparison of the active components and antioxidant activities of extracts from *Abelmoschus esculentus* L. *Pharmagnosy Magazine*, 8(30), 156–161. <http://dx.doi.org/10.4103/0973-1296.96570>
- Lim T. K. (2012). *Edible Medicinal And Non-Medicinal Plants*: Vol 3, Fruits, pp. 160. Springer Science+Business Media B.V. http://dx.doi.org/10.1007/978-94-007-2534-8_21
- Limon-Pacheco, J., & Gonsebatt, M. E. (2009). The role of antioxidants and antioxidant-related enzymes in protective responses to environmentally induced oxidative stress. *Mutation Research*, 6(74), 137–147. <http://dx.doi.org/10.1016/j.mrgentox.2008.09.015>
- Lu, J., Huanfen, L., & Linlin, J. (2011). Chemical constituents in n-butanol extract of *Abelmoschus esculentus*. *Chinese Traditional and Herbal Drugs*, 41, 1771-3.
- Madison, D. (2008). *Renewing America's Food Traditions*. Chelsea Green Publishing. p. 167.
- Mandal, S. M., Roy, A., Mahata, D., Migliolo, L., Nolasco, D. O., & Franco, O. L. (2014). Functional and structural insights on self-assembled nanofiber-based novel antibacterial ointment from antimicrobial peptides, bacitracin and gramicidin S. *The Journal of Antibiotics*, 4 June 2014. <http://dx.doi.org/10.1038/ja.2014.70>
- Maramag, R. P. (2013). Diuretic potential of *Capsicum frutescens* L., *Corchorus olitorius* L., and *Abelmoschus esculentus* L. *Asian journal of natural and applied science*, 2 (1). 60-69.
- Martin, F. (1982). Okra, Potential Multiple-Purpose Crop for the Temperate Zones and Tropics. *Economic Botany*, 36(3), 340–345. <http://dx.doi.org/10.1007/BF02858558>
- Marwat, S. K., Rehman, F. R., Khan, M. A., Ahmed, A., Zafar, M., & Gulam, S. (2011). Medicinal folk recipes used as traditional

- used as traditional phytotherapies in district Dera, Ismail Khan, KPK, Pakistan. *Journal of Botany*, 43(3), 1453-1462.
- Messing, J., Thöle, C., Niehues, M., Shevtsova, A., Glocker, E., ... Hensel, A. (2014). Antiadhesive properties of *Abelmoschus esculentus* (Okra) immature fruit extract against *Helicobacter pylori* adhesion. *PLoS One*, 9(1), e84836. <http://dx.doi.org/10.1371/journal.pone.0084836>
- National Research Council (2006). "Okra, Lost Crops of Africa: Volume II: Vegetables. National Academies Press. ISBN 978-0-309-10333-6. Retrieved 2008-07-15.
- Ndunguru, J., & Rajabu, A. C. (2004). Effect of okra mosaic virus disease on the above-ground morphological yield components of okra in Tanzania. *Scientia Horticulturae*, 99, 225-235 [http://dx.doi.org/10.1016/S0304-4238\(03\)00108-0](http://dx.doi.org/10.1016/S0304-4238(03)00108-0)
- Newsholme, P., Homem D, Bittencourt, P. I., Hagan, C. O., De V, Murphy, C. & Krause, M. S. (2009). Exercise and possible molecular mechanisms of protection from vascular disease and diabetes: the central role of ROS and nitric oxide. *Clinical Science*, 118, 341-349. <http://dx.doi.org/10.1042/CS20090433>
- Newsholme, P., Keane, D., Welters, H. J., & Morgan, N. G. (2007). Life and death decisions of the pancreatic beta-cell: the role of fatty acids. *Clinical Science*, 112, 27-42. <http://dx.doi.org/10.1042/CS20060115>
- Newsholme, P., Rebelato, E., Abdulkader, F., Krause, M., Carpinelli, A., & Curi, R. (2012). Reactive oxygen and nitrogen species generation, antioxidant defenses, and β -cell function: a critical role for amino acids. *Journal of Endocrinology*. 214 (1), 11-20. <http://dx.doi.org/10.1530/JOE-12-0072>
- Ngoc, T. H., Ngo, Q. N., Van, A. T., & Phung, N. V. (2008). Hypolipidemic effect of extracts from *Abelmoschus esculentus* L. (Malvaceae) on Tyloxapol-induced hyperlipidemia in mice. *Warasan Pheatchasat*, 35, 42-46.
- Nwachukwu, E. C, Nulit, R., & Rusea, Go. (2014). Nutritional and biochemical properties of Malaysian okra variety. *Advancement in Medicinal Plant Research*, 2(1), 16-19.
- Odedra, & Nathabhai, K. (2009). Ethnobotany of Maher Tribe In Porbandar District, Gujarat, India. Thesis PhD, Saurashtra University.
- Ofoefule, S. I, & Chukwu, A. (2001). Application of *Abelmoschus esculentus* gum as a mini-matrix for furosemide and diclofenac sodium tablets. *Indian Journal of Pharmaceutical Sciences*, 63 (6), 532-535.
- Oyelade, O. J., Ade-Omowaye, B. I. O., & Adeomi, V. F. (2003). Influence of variety on protein, fat contents and some physical characteristics of okra seeds. *Journal of Food Engineering*, 57, 111-114. [http://dx.doi.org/10.1016/S0260-8774\(02\)00279-0](http://dx.doi.org/10.1016/S0260-8774(02)00279-0)
- Oyenuga, V. A. (1969). *Nigeria's foods and foodstuffs: Their chemistry and nutritive values* (3rd ed.). Ibadan, Nigeria: Ibadan University Press.
- Rajkumari, A., Sarma, K. A., Ilango K. B., Devi, S. D., & Rajak, P. (2012). Studies on the development of colon specific drug delivery system of ibuprofen using polysaccharide extracted from *Abelmoschus esculentus* L. (Moench.) *Asian Journal of Pharmaceutical Sciences*, 7, 67-74.
- Ramachandran, S., Sandeep, V. S, Srinivas, N. K., & Dhanaraju, M. D. (2010). Anti-diabetic activity of *Abelmoschus esculentus* Linn on alloxan-induced diabetic rats. *Research & Reviews in BioSciences*, 4.
- Rao, P. U. (1985) Chemical composition and biological evaluation of okra (*Hibiscus esculentus*) seeds and their kernels. *Plant Foods for Human Nutrition* 35, 389-396. <http://dx.doi.org/10.1007/BF01091784>
- Roy, A, Franco, O. L., & Mandal, S. M. (2013b). Biomedical exploitation of self assembled peptide based nanostructures. *Current Protein and Peptide Science*, 4(7), 580-587. <http://dx.doi.org/10.2174/1389203711209070687>
- Roy, A., Khanra, N., Mishra, A., & Bhattacharyya, N. (2012b). General analysis and Antioxidant study of Traditional fermented drink Handia, its concentrate and volatiles. *Advances in Life Science and its Applications*. 2012, 1, 54-57.
- Roy, A., Khanra, N., Mishra, A., Bhattacharya, C., & Bhattacharyya, N. (2012c). Bakhar-Handia Fermentation: General Analysis and a Correlation between Traditional Claims and Scientific Evidences *Advances in BioResearch*, 3(3), 28.
- Roy, A., Khanra, N., Saha. S., Bhattacharya, C., Mishra, A., & Bhattacharyya, N. (2012a). An antioxidant-rich fermented substrate produced by a newly isolated bacterium showing antimicrobial property against human pathogen, may be a potent nutraceutical in the near future. *Advances in Life Science and its Applications*, 1, 36-44.
- Roy, A., Mahata, D., Paul, D., Korpole, S., Franco, O. L., Mandal, S. M. (2013a) Purification, biochemical characterization and self-assembled structure of a fengycin-like antifungal peptide from *Bacillus thuringiensis* strain SM1. *Frontiers in Microbiology* 21(4), 332. <http://dx.doi.org/10.3389/fmicb.2013.00332>
- Sabitha, V., Ramachandran, S., Naveen, K. R, & Panneerselvam, K. (2012). Investigation of *in vivo* antioxidant property of *Abelmoschus esculentus* (L) moench. fruit seed and peel powders in streptozotocin-induced diabetic rats. *Journal of Ayurveda and Integrative Medicine* 3(4), 188-93. <http://dx.doi.org/10.4103/0975-9476.104432>
- Sabitha, V., Ramachandran, S., Naveen, K. R., & Panneerselvam, K. (2013). Antidiabetic and antihyperlipidemic potential of *Abelmoschus esculentus* (L.) Moench. in streptozotocin-induced diabetic rats. *Journal of Pharmacy and Bioallied Sciences*, 3(3), 397-402. <http://dx.doi.org/10.4103/0975-7406.84447>
- Saha, D., Jain, B, & Jain, V. K. (2011). Phytochemical evaluation and characterization of hypoglycemic activity of various extracts of *Abelmoschus esculentus* Linn. fruit. *International Journal of Pharmacy and Pharmaceutical Sciences*, 3, 183-5.
- Sánchez-Moreno, C., Larrauri, J. A., & Saura-Calixto, F. (1999). Free radical scavenging capacity and inhibition of lipid oxidation of wines, grape juices and related polyphenolic constituents. *Food Research International*, 32, 407-412. [http://dx.doi.org/10.1016/S0963-9969\(99\)00097-6](http://dx.doi.org/10.1016/S0963-9969(99)00097-6)
- Sayana, S. B., Khanwelkar, C. K., Nimmagadda, V. R, Dasi, J. M. B., Chavan, V. R., Kutani, A., & Kotagiri, K. (2014). Evaluation of Diuretic Activity of Alcoholic Extract of Roots of *Cissampelos Pareira* in Albino Rats. *Journal of Clinical and Diagnostic Research*. 8 (5), HC01-HC04.
- Sharma, N., Kulkarni, G. T. & Sharma, A. (2013). Development of *Abelmoschus esculentus* (Okra)-Based Mucoadhesive Gel for

- Nasal Delivery of Rizatriptan Benzoate. *Tropical Journal of Pharmaceutical Research*, 12(2), 149-153.
- Sharma, N., Kulkarni, G. T., Sharma, A., Bhatnagar, A., & Kumar, N. (2013). Natural mucoadhesive microspheres of *Abelmoschus esculentus* polysaccharide as a new carrier for nasal drug delivery. *Journal of Microencapsulation*, 30(6), 589-98. <http://dx.doi.org/10.3109/02652048.2013.764941>
- Shui, G., & Peng L. L. (2004). An improved method for the analysis of major antioxidants of *Hibiscus esculentus* Linn. *Journal of Chromatography A*, 1048, pp. 17-24.
- Siemonsma, J. S. & Kouame, C. (2000). *Abelmoschus esculentus* (L.) Moench. Wageningen Agricultural University, Wageningen, Netherlands.
- Smit, R., Neeraj, K., & Preeti, K. (2013). Traditional Medicinal Plants Used for the Treatment of Diabetes, *International Journal of Pharmaceutical and Phytopharmacological Research*, 3(3), 171-175.
- Steyn, N. P., McHiza, Z., Hill, J., Davids, Y. D., Venter, I., Hinrichsen, E., Opperman, M., Rumbelow, J., & Jacobs, P. (2014). Nutritional contribution of street foods to the diet of people in developing countries: a systematic review. *Public Health Nutrition*, 17(6), 1363-1367. <http://dx.doi.org/10.1017/S1368980013001158> PMID:23680029
- Subrahmanyam, G. V., Sushma, M., Alekya, A., Neeraja, C. H., Harsha, H. S., & Ravindra, J. (2011). Antidiabetic activity of *Abelmoschus esculentus* fruit extract. *International Journal of Research in Pharmacy and Chemistry*, 1, 17-20.
- Sunilson, J. A. J., Jayaraj, P., Mohan M. S., Kumari, A. A. G., Varatharajan, R. (2008). Antioxidant and hepatoprotective effect of the roots of *Hibiscus esculentus* Linn. *International Journal of Green Pharmacy*, 2(4), 200-203.
- Thanakosai, W., & Phuwapraisirisan, P. (2013). First identification of α -glucosidase inhibitors from okra (*Abelmoschus esculentus*) seeds. *Natural Product Communications*. 8 (8), 1085-8.
- Tindall, H. D. (1986). *Vegetables in the Tropics A textbook*. H. D. Tindall (Ed), pp: 328.
- Tomoda, M., Shimizu, N., & Gonda, R. (1985). Isolation and characterisation of a mucilage 'Okra Mucilage R' from the roots of *Abelmoschus esculentus*. *Chemical & Pharmaceutical Bulletin* 33(8), 3330-3335. <http://dx.doi.org/10.1248/cpb.33.3330>
- Tomoda, M., Shimizu, N., Oshima, Y., Takahashi, M., Murakami, M., & Hikino, H. (1987). Hypoglycaemic activity of twenty plant mucilages and three modified products-1. *Planta Medica*, 53(1), 8-12. <http://dx.doi.org/10.1055/s-2006-962605> PMID:3575513
- Tomoda, M., Shimizu, N., Gonda, R., Kanari, M., Yamada, H., & Hikino, H. (1989). Anticomplementary and hypoglycemic activity of okra and hibiscus mucilages. *Carbohydrate Research*. 190(2), 323-8. [http://dx.doi.org/10.1016/0008-6215\(89\)84136-9](http://dx.doi.org/10.1016/0008-6215(89)84136-9)
- Trinh, H. N., Nguyen, N. Q., Tran, T. V. A., & Nguyen V. P. (2008) Hypolipidemic effect of extracts from *Abelmoschus esculentus* L. - Malvaceae on tyloxapol- induced hyperlipidemia in mice. *Mahidol University Journal of Pharmaceutical Science* 35(1-4), 42-46.
- U.S. Department of Agriculture, Agricultural Research Service. (2010). USDA National Nutrient Database for Standard Reference, Release 23. Nutrient Data Laboratory Home Page, <http://www.ars.usda.gov/ba/bhnrc/ndl>
- Uraku, A. J., Ajah, P. M., Okak, A. N., Ibiam, U. A., & Onu, P. N. (2010). Effects of crude extracts of *Abelmoschus esculentus* on albumin and total bilirubin of diabetic albino rats. *International Journal of Science and Nature*. 1, 38-41.
- Uraku, A. J., Onuoha, S. C., Offor, C. E., Ogbanshi, M. E., & Ndidi, U. S. (2011). The effects of *Abelmoschus esculentus* fruits on ALP, AST and ALT of diabetic albino rats. *International Journal of Science and Nature*. 2, 582-6.
- Vaidya, M. V., & Nanoti, M. V. (1989). Bhindi seed powder as coagulant in removal of turbidity from water. *Indian Journal of Environmental Health*, 31(1), 43-48.
- Watt, G, Sir. (1908). *The Commercial Products of India*, p. 381.
- Yesilada, E., Honda, G., Sezik, E., Tabata, M., Fusita, T. ... Takenda, Y. (1995). Traditional Medicine in Turkey, Folk medicine in the inner Taurus mountain. *Journal of Ethnopharmacology*, 46(3), 133-52. [http://dx.doi.org/10.1016/0378-8741\(95\)01241-5](http://dx.doi.org/10.1016/0378-8741(95)01241-5)
- Zaharuddin, N. D., Noordien, M. I, & Kadivar, Ali. (2014). The Use of *Hibiscus esculentus* (Okra) Gum in Sustaining the Release of Propranolol Hydrochloride in a Solid Oral Dosage Form. *BioMed Research International*, Article ID 735891, 8 pages, 2014. <http://dx.doi.org/10.1155/2014/735891>
- Zhou, Y., Zhang, A., Sun, H., Yan, G., & Wang, X. (2014). Plant-derived natural products as leads to antitumor drugs. *Plant Science Today*, 1(2), 46-61. <http://dx.doi.org/10.14719/pst.2014.1.2.17>.

